

Scientific Letter

Cytomegalovirus Genotype Distribution among Postnatally Infected Infants: Association of Glycoprotein B, Glycoprotein N and Glycoprotein H Types with CMV-Associated Thrombocytopenia

Keywords: CMV- associated thrombocytopenia; Glycoprotein B; Glycoprotein N; Glycoprotein H.

Published: September 1, 2020

Received: May 25, 2020

Accepted: August 4, 2020

Citation: Hu H., Cheng Y., Peng Q., Chen K. Cytomegalovirus genotype distribution among postnatally infected infants: association of glycoprotein B, glycoprotein N and glycoprotein H types with CMV-associated thrombocytopenia. Mediterr J Hematol Infect Dis 2020, 12(1): e2020057, DOI: <u>http://dx.doi.org/10.4084/MJHID.2020.057</u>

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<u>https://creativecommons.org/licenses/by-nc/4.0</u>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

To the editor.

Genotyping of CMV has mainly focused on gB, gN, and gH, which play a role in virus entry and may influence the infectivity or pathogenicity of CMV.^{1,2} It has been hypothesized that genetic variation among CMV strains may underlie strain-specific clinical manifestations. Our previous research revealed that there might be a potential association between the genotypes of CMV and neonatal thrombocytopenia, and the detection of some specific genotypes might be indicative of severe manifestations in infants with CMV infection.^{3,4} However, the study design and the criteria to define the study population (congenital and noncongenital cases) and the setting of the control group (CMV-associated thrombocytopenia and nonthrombocytopenic cases) were not clearly established. For this reason, we included patients classified on more unambiguous criteria, and the clinical data collected were complete and thoroughly detailed, which allows us to assess the association between genotypes and the outcome in the non-congenital population.

Methods

Definition. Symptomatic perinatal infection is defined as an infant presenting CMV associated symptoms and positive CMV detection in 3-12 weeks after birth. Symptomatic postnatal infection is referred to as an infant presenting CMV associated symptoms and positive CMV detection after 12 weeks of birth.⁵ Altogether, in the present study, both of them referred to as CMV symptomatic postnatal infection. Moderately to severely symptomatic CMV disease is defined as multiple manifestations attributable to CMV infection. Mildly symptomatic CMV disease is characterized as by one or two isolated features of CMV infection that are mild and transient (e.g., mild hepatomegaly or a single measurement of low platelet count or raised levels of alanine aminotransferase).⁶ Patients. Thirty immunocompetent patients (median, two months; range, 25 days-11 months) with CMVassociated thrombocytopenia were analyzed, including 18 perinatal infections and 12 postnatal infections. Of these 30 patients, 20 were diagnosed with moderately to severely symptomatic CMV disease, and 10 were diagnosed with mildly symptomatic CMV disease. The clinical records of the 30 postnatally infected infants are summarized in Table 1. A group of 40 nonthrombocytopenic individuals, including 20 asymptomatic infants (median, two months; range, 25 days-10 months) and 20 patients (median, two months; range, 29 days–11 months) in CMV infections involving organ systems other than the hematopoietic system from the same period was also included in the study. Among non-thrombocytopenic 20 patients, respiratory symptoms including upper respiratory tract infection (20.0%, 4/20), bronchitis (25.0%, 5/20), and pneumonia (30.0%, 6/20) were the most common symptom at presentation. Other presentations were hepatitis (10.0%). 2/20), jaundice (25.0%, 5/20), and 1 case (5.0%, 1/20) had cholestasis. The baseline characteristics and clinical manifestations in these infants have been described in Table 1 and Table 2.

Laboratory test for CMV infection. Patients were tested for CMV infection using serological CMV tests (IgM and IgG), viral culture, and real-time PCR for blood or urine samples. CMV IgM and CMV IgG were tested using an ELISA kit according to the manufacturer's instructions (DiaSorin S.p.A., Italy). For testing CMV in urine, urine samples were collected and cultured using the shell vial culture method (Chemicon, Temecula, CA, USA). According to the manufacturer's instructions (Daan Gene Company of Zhongshan University, China), fluorescence quantitative CMV-DNA kit was used to quantify of CMV-DNA. DNA level > 10³ copies/ml indicated replication, which was considered positive

| | Gender | Age | | Severity | Mildly: one or two isolated manifestations | | Genotype | | | | |
|-----|--------|-----|--|-------------------|--|-------------------------|----------|-----|---------|--|--|
| No. | | | Other clinical manifestation | of CMV disease | Moderately to severely: multiple manifestations | - Onset of infection | gB | gN | gH | | |
| 1. | F | 2m | - | Mildly | | Perinatal | gB3 | gN3 | gH2 | | |
| 2. | М | 2m | Bronchitis | Moderately | to severely | Perinatal | gB1 | gN4 | gH1+gH2 | | |
| 3. | F | 2m | Jaundice | Moderately | to severely | Perinatal | gB1 | gN1 | gH2 | | |
| 4. | М | 1m | Hepatitis | Moderately | to severely | Perinatal | gB1 | gN4 | gH1 | | |
| 5. | М | 1m | Pneumonia, Anemia | Moderately | to severely | Perinatal | gB1 | gN4 | gH2 | | |
| 6. | М | 2m | Bronchitis, Neutropenia, Anemia | Moderately | to severely | Perinatal | gB1 | gN4 | gH1 | | |
| 7. | М | 1m | Bronchitis, Cholestasis | Moderately | to severely | Perinatal | gB3 | gN4 | gH1 | | |
| 8. | F | 2m | - | Mildly | - | Perinatal | gB1 | gN1 | gH2 | | |
| 9. | F | 1m | Upper respiratory tract infection | Moderately | to severely | Perinatal | gB3 | gN4 | gH2 | | |
| 10. | М | 25d | Jaundice, Neutropenia | Moderately | to severely | Perinatal | gB1 | gN2 | gH2 | | |
| 11. | F | 25d | Jaundice, Neutropenia | Moderately | to severely | Perinatal | gB1 | gN3 | gH2 | | |
| 12. | М | 2m | Neutropenia | Mildly | - | Perinatal | gB2 | gN4 | gH2 | | |
| 13. | М | 29d | Jaundice | Moderately | to severely | Perinatal | gB1 | gN4 | gH1 | | |
| 14. | М | 1m | Anemia | Moderately | to severely | Perinatal | gB1 | gN2 | gH2 | | |
| 15. | М | 2m | Anemia | Moderately | to severely | Perinatal | gB2 | gN2 | gH1 | | |
| 16. | М | 1m | Anemia | Moderately | to severely | Perinatal | gB1 | gN3 | gH1 | | |
| 17. | F | 1m | Anemia, Hepatitis | Moderately | to severely | Perinatal | gB1 | gN2 | gH2 | | |
| 18. | F | 2m | Anemia,Gastrointestinal hemorrhage | Moderately | to severely | Perinatal | gB1 | gN2 | gH1 | | |
| 19. | F | 10m | Upper respiratory tract infection | Mildly | | Postnatal | gB2 | gN2 | gH1 | | |
| 20. | М | 7m | Bronchitis,Fever | | to severely | Postnatal | gB1 | gN4 | gH2 | | |
| 21. | F | 3m | Fever | Mildly | | Postnatal | gB3 | gN1 | gH2 | | |
| 22. | М | 9m | Neutropenia | Mildly | | Postnatal | gB2 | gN4 | gH2 | | |
| 23. | М | 8m | - | Mildly | | Postnatal | gB1 | gN1 | gH1 | | |
| 24. | F | 5m | - | Mildly | | Postnatal | gB1+gB3 | gN4 | gH2 | | |
| 25. | М | 9m | - | Mildly | | Postnatal | gB2 | gN3 | gH2 | | |
| 26. | М | 5m | Neutropenia | Mildly | | Postnatal | gB1 | gN3 | gH1 | | |
| 27. | М | 6m | Pneumonia, Anemia | Moderately | to severely | Postnatal | gB1 | gN4 | gH1 | | |
| 28. | F | 4m | Pneumonia, Anemia, Hepatitis, Neutropenia | Moderately | to severely | Postnatal | gB1 | gN4 | gH1+gH2 | | |
| 29. | М | 11m | Pneumonia, Anemia | Moderately | to severely | Postnatal | gB2 | gN2 | gH2 | | |
| 30. | F | 5m | Upper respiratory tract infection, Fever | Moderately | to severely | Postnatal | gB1 | gN2 | gH2 | | |

in this study. CMV gB, gN and gH genotype analysis was done by nested PCR and restriction length polymorphism as reported.⁷⁻⁹

Statistical analyses. Statistical analysis was conducted using the SPSS ver. 21.0 software (SPSS, Inc., Chicago, IL, USA). Genotype distribution among postnatally infected patients, the relationship between the gB, gN, and gH genotypes and the severity of CMV infections were analyzed using the chi-square test for ratio comparison. Logistic regression analysis was used to assess the associated risk between particular genotypes and the variables of the study. A P-value of less than 0.05 was considered to be statistically significant.

Results

CMV Genotyping. The distribution of gB genotypes in this present study was gB1 (63.3%, 19/30), followed by

gB2 (20.0%, 6/30) and gB3 (13.3%, 4/30). We also found 1 coinfection case (3.3%, 1/30) with 2 genotypes (gB1/gB3), no gB4 genotype was found. Notably, significantly higher frequency of gB1 (80.0%,16/20) was found in moderately to severely CMV infection infants compared to infants with mildly symptomatic

CMV disease ($\chi 2= 8.132$, p = 0.043) (Figure 1).

The overall distribution of individual genotypes in this study cohort was as follows: gN1(13.3%,4/30), gN2(26.7%,8/30), gN3 (16.7%,5/30) and gN4(43.3%,13/30). Comparing distribution in 20 asymptomatic infants with CMV infection, the gN1(5.0%,1/20) was the less prevalent genomic variants in moderately to severely CMV infection patients ($x^2-15.007$, n=0.002) (Figure 1)

 $(\chi 2=15.097, p=0.002)$ (Figure 1).

The gH1, gH2 and gH1/gH2 genotypes were distributed in 36.7% (11/30), 56.7% (17/30) and 6.7%

| Asymptomatic CMV infection infants | | | | | | | Non-thrombocytopenic patients | | | | | | | | |
|------------------------------------|--------|-----|----------|-----|-----|--------|-------------------------------|------------------------|-----------------------------------|-----------|-----|-----|-----|--|--|
| No. | Gender | Age | Genotype | | No. | Gender | 1 00 | Clinical manifestation | Onset of | Genotype | | | | | |
| | | | gB | gN | gH | INO. | Gender | Age | Clinical manifestation | infection | gB | gN | gH | | |
| 1. | F | 2m | gB2 | gN1 | gH2 | 1. | F | 3m | Upper respiratory tract infection | Postnatal | gB3 | gN1 | gH1 | | |
| 2. | М | 1m | gB1 | gN1 | gH1 | 2. | М | 2m | Bronchitis | Perinatal | gB1 | gN4 | gH1 | | |
| 3. | F | 1m | gB3 | gN4 | gH1 | 3. | М | 2m | Pneumonia | Perinatal | gB3 | gN1 | gH1 | | |
| 4. | F | 28d | gB1 | gN3 | gH2 | 4. | F | 1m | Hepatitis, Jaundice | Perinatal | gB2 | gN1 | gH1 | | |
| 5. | М | 2m | gB3 | gN3 | gH1 | 5. | М | 4m | Pneumonia | Postnatal | gB2 | gN3 | gH2 | | |
| 6. | М | 2m | gB1 | gN2 | gH1 | 6. | F | 5m | Upper respiratory tract infection | Postnatal | gB1 | gN4 | gH1 | | |
| 7. | М | 1m | gB2 | gN1 | gH1 | 7. | М | 2m | Jaundice | Perinatal | gB1 | gN4 | gH1 | | |
| 8. | М | 2m | gB2 | gN3 | gH2 | 8. | F | 2m | Pneumonia | Perinatal | gB1 | gN4 | gH1 | | |
| 9. | F | 25d | gB1 | gN3 | gH1 | 9. | М | 3m | Pneumonia | Postnatal | gB1 | gN4 | gH2 | | |
| 10. | М | 1m | gB1 | gN3 | gH2 | 10. | F | 5m | Bronchitis | Postnatal | gB1 | gN3 | gH2 | | |
| 11. | F | 2m | gB3 | gN1 | gH1 | 11. | F | 29d | Hepatitis, Jaundice | Perinatal | gB1 | gN4 | gH1 | | |
| 12. | F | 2m | gB2 | gN4 | gH2 | 12. | F | 5m | Bronchitis | Postnatal | gB2 | gN4 | gH1 | | |
| 13. | М | 2m | gB2 | gN4 | gH1 | 13. | F | 7m | Upper respiratory tract infection | Postnatal | gB1 | gN4 | gH1 | | |
| 14. | F | 4m | gB3 | gN3 | gH1 | 14. | М | 10m | Upper respiratory tract infection | Postnatal | gB1 | gN4 | gH2 | | |
| 15. | М | 6m | gB1 | gN3 | gH2 | 15. | F | 2m | Pneumonia | Perinatal | gB3 | gN2 | gH2 | | |
| 16. | М | 10m | gB1 | gN1 | gH2 | 16. | М | 29d | Jaundice | Perinatal | gB1 | gN4 | gH2 | | |
| 17. | F | 3m | gB1 | gN3 | gH1 | 17. | М | 1m | Jaundice, Cholestasis | Perinatal | gB1 | gN1 | gH1 | | |
| 18. | F | 5m | gB1 | gN4 | gH1 | 18. | М | 2m | Bronchitis | Perinatal | gB2 | gN4 | gH2 | | |
| 19. | F | 5m | gB3 | gN3 | gH1 | 19. | F | 2m | Bronchitis | Perinatal | gB3 | gN2 | gH1 | | |
| 20. | М | 9m | gB1 | gN1 | gH1 | 20. | F | 11m | Pneumonia | Postnatal | gB1 | gN4 | gH2 | | |

Table 2. The baseline characteristics and distribution of CMV genotypes among non-thrombocytopenic infants.

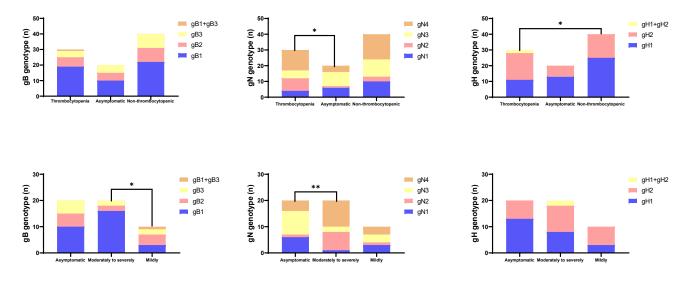


Figure 1. Distribution of CMV genotypes in different groups. *, p < 0.05; **, p < 0.01

(2/30) of the patients, respectively (Figure 1). Compared with the genotype distribution in non-thrombocytopenic infants, a greater frequency of gH2 in CMV-associated thrombocytopenia infants was noted with significant difference ($\chi 2=6.269$, p = 0.044). No difference in the distribution of gH genotypes in symptomatic and asymptomatic patients, or in moderately to severely symptomatic CMV disease and mildly symptomatic CMV disease (Figure 1).

Genotype Association With CMV-associated thrombocytopenia and severity of CMV disease. In the logistic regression analysis, the gN2 [p = 0.043, with OR=4.598, 95%CI (1.052-20.098)] and gH2 [p = 0.038, with OR=2.933, 95%CI (1.060-8.117)] genotypes were associated with an elevated risk of developing thrombocytopenia. Besides, gB1 [p = 0.022, with OR=9.820, 95%CI (1.400-68.888)] represented the most virulent genotypes and was associated with severe manifestations in CMV-associated thrombocytopenia infants. Conversely, the gN1 [p = 0.044, with OR=0.061, 95%CI (0.004-0.930)] genotype was associated with a reduced risk of severely symptomatic CMV disease.

Discussion. The gB of CMV likely plays a crucial role in viral entry into cells, the transmission of the virus from cell to cell, and the fusion of infected cells. It has been reported that the gB genotypes vary in their ability to stimulate cell-mediated or cytotoxic immune response.^{10,11} Therefore, variations in gB are likely to have significant effects on the pathogenesis of CMV disease and the spectrum of host cells infected by the virus. Our previous studies also confirmed that the gB1 genotype had more virulence in infants with symptomatic CMV disease.^{3,4}

But interestingly, in asymptomatic infected infants, gB1 was also the dominant genotype, and its genotype distribution was not significantly different from that of CMV-associated thrombocytopenia infants.

Consequently, we speculate that CMV gB1 strains may elicit a severe immunopathological response that in some infants can control the symptoms of CMV and, in others, lead to CMV-associated thrombocytopenia with organ damage and disease manifestations. However, the virulence of gB1 in asymptomatic infants is negligible in relationship with a difference in the individual immune status.

The CMV strain with gN1 genotype may represent a less virulent virus phenotype, especially considering that the variation is a typical AD169-like glycoprotein, which is far away from CMV clinical isolates in immunology.¹²⁻¹⁴ In our study, among CMV-associated thrombocytopenia infants (20 cases) who were classified as having moderately to severely symptomatic CMV disease, 17 had gN4 or gN2 genotypes and only one had a gN1 genotype, supporting the idea that gN1 genotype may be less virulent. In addition, compared with the genotype distribution in asymptomatic and non-

thrombocytopenic infants in present study, thrombocytopenia occurred more frequently in infants infected with the CMV gN2 genotype, although the proportion of this genotype was less than that of gN4 in CMV-associated thrombocytopenia infants. The gN2 genotype was detected in 26.7% (8/30) of infants with CMV-associated thrombocytopenia and was associated with at least a 4-fold increased risk of developing thrombocytopenia. Our study is the first to demonstrate that a gN variant might be associated with a risk of CMV-associated thrombocytopenia in infants infected postnatally.

As we reported earlier, the gH2 genotype was associated with at least a 7-fold increased risk of developing CMV-associated thrombocytopenia among infants with congenital and perinatal infections⁴. After including postnatal infection and non-thrombocytopenic cases into the analysis, similar conclusions were reached.

Based on these cases, several general points can be highlighted. First, in regression analysis, the difference in the setting of the non-thrombocytopenic control group, which includes asymptomatic and symptomatic infants, may cause a discrepancy in results. Increasing the sample size and choosing an appropriate scale setting may reduce this discrepancy. Second, a specific cytomegalovirus genotype may show strong virulence in some CMV- related diseases, while in other CMVrelated diseases or asymptomatic infants, it may not show corresponding characteristics of virulence. Finally, in addition to CMV gB, gN, and gH, CMV glycoprotein also includes gO, gM and gL. Six glycoproteins are essential for fibroblasts to enter CMV, and form glycoprotein complexes, gCI (gB), gCII (gM / gN), gcIII (gH / gL / gO) on the virus membrane.¹⁵ In the study of a CMV- related disease, it is more reasonable to include all essential CMV glycoprotein genotypes into the analysis.

Hongbo Hu¹, Ying Cheng², Qiaoying Peng³ and Kun Chen⁴.

¹ Department of Laboratory, Maternal and Child Health Hospital of Hubei Province, China.

- ² Department of Pediatrics, Maternal and Child Health Hospital of Hubei Province, China.
- ³ Department of Neonatology, Maternal and Child Health Hospital of Hubei Province, China.

⁴ Department of Laboratory, Wuhan Ninth Hospital, China.

Competing interests: The authors declare no conflict of Interest.

Correspondence to: Kun Chen, Department of Laboratory, Wuhan Ninth Hospital, No. 20, Jilin Street, Qingshan District, Wuhan 430081, China. Tel: 86-027-68865331. E-mail: <u>chenkun430922@163.com</u>

References:

- Ross SA, Pati P, Jensen TL, et al. Cytomegalovirus genetic diversity following primary infection. J Infect Dis. 2020;221(5):715-720. <u>https://doi.org/10.1093/infdis/jiz507</u> PMid:31593588
- 2. Nahar S, Hokama A, Iraha A, et al. distribution of cytomegalovirus genotypes among ulcerative colitis patients in Okinawa, Japan. Intest Res.

2018 ;16(1):90-98. https://doi.org/10.5217/ir.2018.16.1.90 PMid:29422803 PMCid:PMC5797277

 Hu H, Cheng Y, Peng Q, Chen K. Clinical Features, Treatment Courses, and Distribution of Cytomegalovirus Genotypes among Thrombocytopenia Patients Aged Younger than 12 Months [published online ahead of print, 2020 Jun 11]. Am J Perinatol. 2020;10. https://doi.org/10.1055/s-0040-1713001

- Hu H, Peng W, Peng Q, Cheng Y. Cytomegalovirus Genotype Distribution among Congenital and Perinatal Infected Patients with CMV-Associated thrombocytopenia [published online ahead of print, 2020 Jun 1]. Fetal Pediatr Pathol. 2020;1-10. <u>https://doi.org/10.1080/15513815.2020.1765916</u> PMid:32479132
- Shen Z, Shang SQ, Zou CC, Zheng JY, Yu ZS. The detection and clinical features of human cytomegalovirus infection in infants. Fetal Pediatr Pathol. 2010;29(6):393-400. <u>https://doi.org/10.3109/15513815.2010.494705</u> PMid:21043563
- Rawlinson WD, Boppana SB, Fowler KB, et al. Congenital cytomegalovirus infection in pregnancy and the neonate: consensus recommendations for prevention, diagnosis, and therapy. Lancet Infect Dis. 2017;17(6): e177-e188. https://doi.org/10.1016/S1473-3099(17)30143-3
- Yu ZS, Zou CC, Zheng JY, Zhao ZY. Cytomegalovirus gB genotype and clinical features in Chinese infants with congenital infections. Intervirology. 2006;49(5):281-285.
 <u>https://doi.org/10.1159/000093458</u> PMid:16714857
- Guo S, Yu MM, Li G, Zhou H, Fang F, Shu SN. Studies on genotype of human cytomegalovirus glycoprotein H from infantile clinical isolates. Zhonghua Er Ke Za Zhi. 2013;51(4):260-264 (in Chinese).
- Chen HP, Lin JC, Yang SP, et al. The type-2 variant of human cytomegalovirus glycoprotein N(gN-2) is not the rarest in the Chinese population of Taiwan: influence of primer design. J Virol Methods.2008;151(1):161-164. https://doi.org/10.1016/j.jviromet.2008.03.018

PMid:18499272

 Saccoccio FM, Jenks JA, Itell HL, et al. Humoral immune correlates for prevention of postnatal cytomegalovirus acquisition. J Infect Dis. 2019;220(5):772-780. <u>https://doi.org/10.1093/infdis/jiz192</u>

PMid:31107951 PMCid:PMC6667799
11. Lee S, Doualeh M, Affandi JS, Makwana N, Irish A, Price P. Functional and clinical consequences of changes to natural killer cell phenotypes driven by chronic cytomegalovirus infections. J Med Virol. 2019;91(6):1120-1127. https://doi.org/10.1002/jmv.25401

 PMid:30636352
 Paradowska E, Jabłońska A, Studzińska M, et al. Distribution of cytomegalovirus gN variants and associated clinical sequelae in infants. J Clin Virol. 2013;58(1):271-275. https://doi.org/10.1016/j.jcv.2013.05.024
 PMid:23806667

- Pignatelli S, Rossini G, Dal Monte P, Gatto MR, Landini MP. Human cytomegalovirus glycoprotein N genotypes in AIDS patients. AIDS. 2003; 28;17(5):761-763. <u>https://doi.org/10.1097/00002030-200303280-00018</u> PMid:12646803
- 14. Mujtaba G, Khurshid A, Sharif S, et al. distribution of cytomegalovirus among neonates born to infected mothers in Islamabad, Pakistan. PLoS One. 2016 ;11(7): e0156049. <u>https://doi.org/10.1371/journal.pone.0156049</u> PMid:27367049 PMCid:PMC4930188
- Coleman S, Hornig J, Maddux S, et al. Viral glycoprotein complex formation, essential function and immunogenicity in the guinea pig model for cytomegalovirus. PLoS One. 2015;12;10(8): e0135567. <u>https://doi.org/10.1371/journal.pone.0135567</u> PMid:26267274 PMCid:PMC4534421