

Review Article

EBV and HIV-Related Lymphoma

Michele Bibas and Andrea Antinori

Clinical Department, National Institute for Infectious Diseases “Lazzaro Spallanzani”, IRCCS, Rome, Italy

Correspondence to: Michele Bibas, MD, Clinical Department, National Institute for Infectious Diseases, “Lazzaro Spallanzani, IRCCS, via Portuense 292 – 00149 Rome, Italy. Phone: +39 06 55170480, Fax: +39 06 55170477. E-mail: michele.bibas@inmi.it

Published: December 29, 2009

Received: December 23, 2009

Accepted: December 27, 2009

Medit J Hemat Infect Dis 2009, 1(2): e2009032 DOI 10.4084/MJHID.2009.032

This article is available from: <http://www.mjhid.org/article/view/5272>

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract: HIV-associated lymphoproliferative disorders represent a heterogeneous group of diseases, arising in the presence of HIV-associated immunodeficiency. The overall prevalence of HIV-associated lymphoma is significantly higher compared to that of the general population and it continues to be relevant even after the wide availability of highly active antiretroviral therapy (HAART) (1). Moreover, they still represent one of the most frequent cause of death in HIV-infected patients. Epstein–Barr virus (EBV), a γ -Herpesviruses, is involved in human lymphomagenesis, particularly in HIV immunocompromised patients. It has been largely implicated in the development of B-cell lymphoproliferative disorders as Burkitt lymphoma (BL), Hodgkin disease (HD), systemic non Hodgkin lymphoma (NHL), primary central nervous system lymphoma (PCNSL), nasopharyngeal carcinoma (NC). Virus-associated lymphomas are becoming of significant concern for the mortality of long-lived HIV immunocompromised patients, and therefore, research of advanced strategies for AIDS-related lymphomas is an important field in cancer chemotherapy. Detailed understanding of the EBV lifecycle and related cancers at the molecular level is required for novel strategies of molecular-targeted cancer chemotherapy. The linkage of HIV-related lymphoma with EBV infection of the tumor clone has several pathogenetic, prognostic and possibly therapeutic implications which are reviewed herein.

Epidemiology of AIDS related lymphomas:

Registry linkage studies in the pre-highly active antiretroviral therapy (HAART) era found that the incidence of high grade B-cell non-Hodgkin's lymphoma (NHL) in HIV-infected individuals was 60-200 times higher than that in HIV-uninfected persons. The introduction of HAART during the mid-1990s has been associated with a fall in

incidence of opportunistic infections and AIDS-associated malignancies, including NHL^{1,2}.

Within the French Hospital Database on HIV Infection (FHDH), the incidence of systemic NHL has decreased between 1993 and 1994 and between 1997 and 1998, from 8.6 per 1,000 to 4.3 per 1,000 person-years, respectively³; the incidence in the same cohort was 2.8 per 1,000 person-years in 2006. This is consistent with reports of decreased

incidence of HIV-related NHL in the post HAART era from the U.K., Australia, California⁴. Nevertheless, the incidence ratio of NHL still remains relatively high in HIV-infected patients (5-6). On the contrary, the incidence of PCNSL has dramatically decreased since the introduction of HAART⁴. Concerning HD, the relative risk is increased, ranging from five- to 25-fold compared to that of the general population^{7,8,9}.

Approximately 1–6% of HIV infected patients develop lymphoma each year. In 2006 the World Health Organization estimated 39.5 million people were living with HIV and that during that year there were 4.3 million new infections with 65% of these occurring in sub-Saharan Africa. Major increases were also seen in Eastern Europe and Central Asia, where it appears that infection rates have risen by more than 50% since 2004. Many of those with retroviral infection will either have limited access to HAART or will be unaware of their HIV status. Therefore the incidence of HIV-associated lymphomas will most likely increase globally in the years to come^{10,11}.

Categories of HIV-associated lymphoma: The WHO (12) classification of lymphoid neoplasms categorises (Table 1) the HIV-associated lymphomas into:

1. Those also occurring in immunocompetent patients, as Burkitt and Burkitt-like lymphomas, Diffuse large B-cell lymphomas, Centroblastic and Immunoblastic (including primary CNS Lymphoma), Extranodal marginal zone lymphoma of Malt Type, Peripheral T-cell lymphoma, Classical Hodgkin lymphoma (80% of all HIV lymphomas);
2. Those occurring more specifically in HIV-positive patients as Primary Effusion Lymphoma⁴ and Plasmablastic Lymphoma of oral cavity type and other variants (3%);
3. Those also occurring in patients with other forms of immunosuppression as Polymorphic B-cell lymphoma (PTLD-like) (5% of all HIV lymphomas).

Immunodeficiency and pathogenesis of lymphomas in HIV-infected individuals: HIV is a lentivirus of the retrovirus family, and thus integrates into host chromosomal DNA using a DNA intermediate. It has been generally believed that integration of HIV is a random process, and therefore this process is not in itself oncogenic¹³.

Table 1. Classification of HIV-associated lymphomas

1. Lymphoma also occurring in immunocompetent patients:
a. Burkitt and Burkitt-like Lymphoma
b. Diffuse large B-cell lymphoma
i. Centroblastic
ii. Immunoblastic (including primary CNS lymphoma)
c. Extranodal marginal zone lymphoma of Malt type
d. Peripheral T-cell lymphoma
e. Classical Hodgkin Lymphoma
2. Lymphoma occurring more specifically in Hiv positive patients
a. Primary effusion Lymphoma
b. Plasmablastic lymphoma of the oral cavity type
3. Lymphoma also occurring in other immunodeficiency states
a. Polymorphic B-cell lymphoma (PTLD-like)

Accordingly with this theory is the fact that Southern blot analysis of HIV-associated lymphomas has failed to detect HIV sequences¹⁴, with rare reports of clonal integration restricted to T-cell neoplasms¹⁵. Although the neoplastic cells are not themselves infected with HIV in most cases, in vitro evidence suggests that HIV does have transforming properties. Laurence and Astrin showed that HIV infection of B-cell lines derived from EBV-seropositive individuals led to B-cell immortalisation, dysregulation of MYC, and activation of EBV⁷. Certain HIV gene products, particularly Tat, have been implicated as potentially oncogenic in their role as transactivators of cellular genes, such as IL6 and IL10⁷. Tat protein can more directly interfere with cell cycle control by interaction with the regulatory protein Rb2/p130⁸. This role of the Tat protein has been proposed as a significant factor in the pathogenesis of HIV-related Burkitt lymphoma⁸.

The predominant contribution of HIV to lymphoma pathogenesis is believed to be through indirect mechanisms. The increased risk for lymphoma among HIV-infected individuals appears related to multiple factors, including duration and degree of immunosuppression, induction of cytokines leading to B-cell proliferation, and opportunistic infections with oncogenic herpesviruses such as EBV and HHV8¹⁴.

HIV-associated malignancies are commonly considered to be the result of diminished immune surveillance against viruses and virus-infected tumor cells. The beneficial effects of HAART on these tumors have therefore been interpreted as the result of drug-mediated HIV suppression and immune reconstitution.

This is supported by several findings. For example, EBV load is increased in patients before development of B-cell lymphoma, whereas specific immune responses against the virus are decreased^{16,17,18}. The relative risk of AIDS-associated malignancies increases progressively as a function of the progressive decline of CD4+ T-cell counts¹⁹. Nevertheless, the relation between immune deficiency and tumor development is not straightforward.

In fact, only certain types of AIDS-associated tumors arise in immunodeficient patients. In particular, NHL subtypes including Immunoblastic lymphomas and PCNSL, along with Burkitt's-like lymphomas, typically develop in patients with very low CD4+ T-cell counts. On the other hand, the incidence of other NHL subtypes such as Centroblastic Diffuse Large-cell Lymphomas, along with classic Burkitt's Lymphoma, Hodgkin's disease, cervical cancer and, most notably, Kaposi's sarcoma, increases in patients who have significantly higher CD4+ T-cell numbers^{9,20,21}.

The overall risk of tumour development is very high in HIV-infected individuals, but the relative increase in tumor risk with stepwise decreases in CD4+ T-cell counts is only marginal²². It has been observed that the risk of tumor development increases steeply as CD4+ T-cell counts decline below a certain threshold, nevertheless, once below this threshold, cancer risk becomes less dependent on further CD4+ T-cell loss¹⁹.

However, evidence indicates that this hypothetical CD4+ T-cell count threshold can be very high in certain individuals. In particular, in HIV-infected homosexual men, the incidence rate of Kaposi's sarcoma increases by more than 1000-fold before a consistent CD4+ T-cell decline²⁰. So, CD4+ T-cell loss and consequent immune deficiency cannot fully explain the increased incidence of certain malignancies in HIV-infected individuals. Indeed, several recent studies show that immune activation causes and precedes the development of immune deficiency in HIV infection^{23,24,25}. Sustained and uncontrolled HIV replication leads to continuous antigenic stimulation and to chronic T-cell activation and proliferation, which, in turn, generates a continuous drain of

naive and memory T cells that become activated, proliferate, die by apoptosis or re-enter the pool of memory T cells. However, this exhausts the pool of naive T cells, impairing the capacity to mount antigen-specific immune responses^{22,23,24,25}.

Several other studies also indicate that immune activation, rather than immune deficiency, is the key factor in the initiation of B-cell lymphomas. In particular, AIDS-associated B-cell lymphomas are described to be preceded by chronic antigen dependent B-cell stimulation leading to a persistent and generalized lymphadenopathy that, in turn, promotes the clonal expansion of pre-neoplastic antigen-specific B-cell populations^{26,27}.

Furthermore, an increased EBV load precedes the development of B-cell lymphoma¹⁷, whereas extracellular Tat increases B-cell proliferation and induces B-cell lymphomas in mice^{26,27}.

The role of EBV: Regarding EBV, the percentage of cases within each histotypes with EBV viral infection is variable, ranging from 60% to 100%. In contrast to other lymphomas, a high frequency of EBV association has been shown in HL (80%-100%) tissues from HIV-infected people and the EBV-transforming protein, EBV-encoded latent membrane protein-1 (LMP-1), is expressed in virtually all HIV-HL cases^{28,29}. On this basis, HL in HIV-infected persons appears to be an EBV-driven lymphoma³⁰.

The spectrum of lymphomas occurring in HIV-infected patients includes pathologic subtypes displaying specific association with distinct viruses. BL and DLBCL-IB with plasmacytoid differentiation are often HIV associated and closely linked to EBV infection.

The HIV-associated DLBCL-IB is distinct from other large cell lymphomas occurring in both HIV-seropositive and -seronegative patients because HIV-associated DLBCL-IB lymphomas display a plasma cell-related phenotype.

Most HIV-associated lymphoproliferative disorders, including primary central nervous system lymphoma, systemic DLBCL IB-plasmacytoid, PEL and its solid variant, and PBLs of the oral cavity type, display a phenotype related to plasma cells and are linked to EBV infection.

Burkitt lymphoma: Among EBV-positive high-grade B cell Lymphomas, Burkitt Lymphoma (BL) occupies a particular position as being the tumor type in which EBV was discovered. Burkitt and Burkitt-like/atypical Burkitt lymphomas make up the largest group of HIV-associated non-Hodgkin

lymphomas, comprising up to 35–50% of these neoplasms in some studies³¹.

Classification of these lymphomas in the HIV setting follows the same diagnostic criteria as are used in the general patient population. That is, a diagnosis of Burkitt or Burkitt-like lymphoma requires a medium-sized CD10-positive B-cell population with a high proliferative rate and demonstration of a translocation involving the MYC gene¹². Peripheral blood involvement is less common in HIV-infected patients compared to HIV-negative patients with Burkitt lymphoma, although it can occur^{12,31,32}. Burkitt lymphoma occurring in the HIV setting is characterised by multiple genetic lesions, with the relative significance of each in the pathogenesis of this lymphoma unknown. In addition to the translocation involving MYC, point mutations in regulatory regions associated with MYC and within the TP53 tumour suppressor gene are common¹².

In the context of HIV infection, EBV-encoded RNA (EBER) can be detected by in situ hybridisation in tumor cells in about 30% of Burkitt lymphomas, 50–70% of Burkitt lymphomas with plasmacytoid differentiation, and 30–50% of Burkitt-like lymphomas.

Similarly to sporadic or epidemic forms of Burkitt lymphoma, in HIV-associated EBER-positive disease the viral oncogenes LMP-1 and EBNA-2 are not expressed (**Table 2, Table 3**).

Although not essential in the pathogenesis of BL, EBV supports tumor development. EBNA-1, a viral protein required for the replication and maintenance of the latent viral episomal DNA, is found consistently in BL cells³³. The presence of latent EBV in BL cells has been shown to promote genetic instability (34), suggesting a mechanism by which latent EBV could contribute to genetic alterations required for the development of BL.

This is in contrast to EBER-positive immunoblastic DLBCL and PEL, which do show expression of these EBV-associated viral oncogenes. Thus EBV may not play the same role in oncogenesis in these different types of lymphoma. It is interesting to note that although Burkitt lymphoma is common in HIV-infected patients, it is not associated with other forms of immunosuppression.

This may indicate that the oncogenic properties of HIV itself play a greater role in pathogenesis in this highly proliferative tumour compared with EBV or that there are other mechanisms. Dysregulation of cell cycle proteins has been implicated in the development of Burkitt

lymphoma. Inactivating mutations of the tumour suppressor gene RBL2 (Rb2/p130) are frequently found in endemic Burkitt lymphoma, and are also found in sporadic cases³⁵.

By contrast, in HIV-associated cases, abnormal overexpression of wild-type RBL2 is seen. This finding, in conjunction with studies indicating that the function of Rb2/p130 in the control of the G0/G1 transition can be negated by physical interaction with the Tat protein of HIV-1, may suggest a direct role for HIV proteins acting synergistically with MYC activation in the pathogenesis of Burkitt lymphoma³⁶.

Diffuse large B-cell Lymphoma: As in the HIV-negative setting, the category of HIV associated DLBCL is a clinically and pathologically heterogeneous group. Lymphomas with a predominance of centroblasts have been termed centroblastic DLBCL, whereas those with greater than 90% immunoblasts/plasmablasts have been termed immunoblastic DLBCL.

These two general morphological subtypes show correlation with certain clinical features and molecular profiles. The subtypes occur with approximate equal frequency in HIV-infected patients, with the relative frequency of centroblastic DLBCL increasing and that of immunoblastic DLBCL decreasing in recent years due to advances in HIV therapy. Centroblastic DLBCL occurs in the setting of mild immunosuppression, has a low frequency of EBV positivity (30–40%) without expression of LMP-1, shows a germinal centre B-cell phenotype (expression of CD10 and BCL6, and lack of expression of CD138 and MUM1), and frequently shows rearrangements of the BCL6 gene.

In contrast, immunoblastic DLBCL usually occurs in the context of severe immunosuppression, has a high frequency of EBV positivity (80–90%) with frequent expression of LMP-1 and EBNA-2, shows a non-germinal centre B-cell/activated B-cell phenotype (lack of expression of CD10 and BCL6, expression of CD138 and MUM1), and lacks rearrangements of BCL6³⁵ (**Table 2, Table 3**).

The transforming EBV protein LMP-1 is frequently expressed^{37,38}. LMP-1 plays a crucial role in the transformation of B-lymphocytes by EBV⁴⁰. Thus, LMP-1 transforms rodent fibroblasts⁴⁰ transgenic mice that express LMP-1 in B cells show increased development of B-cell lymphomas⁴¹ and LMP-1 deletion mutants of EBV are compromised in their ability to immortalize human primary B cells⁴². LMP-1 activates the NFκB as well as the JNK and p38 pathways (39,40,41), by recruiting

Table 2. Immunological and EBV status in AIDS-related Lymphomas

Lymphoma Histology	immunodeficiency	% in HIV	EBV+ Rate	Viral cofactor
Systemic AIDS-Related Lymphomas				
Burkitt Lymphoma	Mild	55		EBV EBER
Classic BL		30	30%	
BL with Plasmocitoid diffent		20	50-70%	
Atypical BL		Less freq	30-50%	
Diffuse Large B-cell lymphoma		30		
Centroblastic type	Mild	20	30-40%	
Immunoblastic type	Marked	10	90-100%	EBV LMP1
AIDS Primary CNS Lymphoma	Marked	< 5	100%	EBV LMP1
Primary effusion Lymphoma	Marked	< 5	90%	EBV
Plasmablastic lymphoma oral cavity		< 5	50%	LMP1
Hodgkin Disease Classical	Marked		100%	LMP1 LMP2A

cellular TRAF 1-3 and TRADD molecules to 2 short sequence motifs, CTAR-1 and CTAR-2, respectively, in the cytoplasmic domain of the LMP-1 molecule^{43,44,45}.

In B cells, LMP-1 increases the expression of the antiapoptotic proteins A20 and bcl-2, the adherence molecule CD54/ICAM-1, the cell-cycle regulator p27Kip,71 and many others⁴⁶.

In DLBCL, expression of LMP-1 correlates inversely with the expression of BCL6, a marker for germinal center B cells, suggesting that, among DLBCLs, the impact of EBV LMP-1 is likely to be strongest in tumors representing a post-germinal center plasmacytic differentiation profile⁴⁷. In addition, knockdown of LMP-1 in cell lines derived from AIDS-DLBCL results in apoptosis, indicating that this viral oncoprotein plays a role in lymphoma pathogenesis⁴⁸.

EBV-associated DLBCLs have t been considered as EBV-driven lymphoproliferations occurring in the context of a defective T-cell immunity against EBV.⁴⁹ However, unlike EBV-driven lymphoproliferative disease in transplant recipients, which includes monoclonal, oligoclonal, as well as polyclonal B-cell proliferations, DLBCL is always monoclonal. This suggests that, in addition to the effects contributed by EBV LMP-1, additional factors such as genetic damage are likely to contribute to the pathogenesis of AIDS-DLBCL.

Primary CNS Lymphoma: Accounting for 15% of HIV-associated lymphomas, PCNSL has a reported incidence of over 1000 times greater than in the non-HIV population⁵⁰. This is most likely a reflection of the brain as a relatively immunoprivileged site. There has been a decline in its incidence since HAART introduction⁵¹, and it would confirm the strong association of this tumor with severe and prolonged immunosuppression. Clinical presentation results from neurological deficits related to the site of the tumor, with mental state disturbance and seizures more common than in non-HIV PCNSL. Systemic B symptoms are also common^{52,53}.

These tumors have a tendency to occur late in the course of HIV infection and show EBV association in virtually 100% of the cases⁵³. A few studies have reported that detection of EBV in the cerebrospinal fluid of HIV-positive patients with a CNS lesion infers a diagnosis of lymphoma^{54,55,56}. These lymphomas have been reported to express all EBV latent encoded proteins (latency III)⁵⁷, and there are observations consistent with their histogenetic derivation from germinal center-related B cells.⁵⁸ Nevertheless, the exact role of EBV in the pathogenesis of these disorders remains not completely defined (**Table 2, Table 3**).

Table 3. Features of EBV-associated Aids-associated B-cell Lymphoma

Lymphoma	EBV %	EBV Latency	Phenotype	Cellular origin of lymphoma cells
Hodgkin L Classical	100%	Type II	Loss B-cell Phenotype	Pre-apoptotic GC B cells
PCNSL	100%	Type III	BCL6- CD138+ Mum 1+	GC or post GC B cells
Burkitt Lymphoma	55%	Type I	BCL6+ CD10+ CD77+	GC B cell
PEL	90-100%	Type I	Loss B-cell Phen. CD38+	GC or post-GC B cells
DLCL-CB	30%	Type I	BCL6+ CD138- MUM1-	Mostly GC or post-GC B cells
DLCL-IB	90%	Type III	BCL6- CD138+MUM1+	Mostly GC or post-GC B cells

Most patients have CD4 counts <50/uL and have multifocal lesions at time of diagnosis. Ocular involvement occurs in up to 20% of cases⁵⁹. Full staging at time of diagnosis is essential to exclude system NHL involving the brain. MRI brain scan has a higher diagnostic yield than CT and is recommended for suspected intracranial masses⁶⁰. Up to 30% of CNS lesions in HIV patients are found to be PCNSL with toxoplasmosis and progressive multifocal leukoencephalopathy making up the remaining cases⁶⁰. The most common histology is immunoblastic variant DLBCL.

Differentiation between PCNSL and toxoplasmosis can be difficult, as both cause ring enhancing lesions with mass effect and oedema (although PCNSL lesions are more likely to be periventricular) and up to 15% false negative rates for toxoplasmosis serology^{61,62}.

Radionuclide scanning has also been investigated. PCNSL lesions are avid by Thallium²⁰¹ single photon emission CT and fluorodeoxyglucose-positron¹⁸ emission tomography (FDG-PET), however improve specificity should be combined with PCR and is emerging as an alternative to brain biopsy^{63,64}. This needs to be further validated and brain biopsy is still the definitive diagnostic procedure, but must be weighed against a mortality rate of 2–3%⁶⁴, particularly in the post-HAART era during which it seems that EBV-DNA detection shows a reduced negative predictive value compared to that of the pre-HAART period.

In the new trials the use of EBV-DNA measurement is used as a surrogate to brain biopsy. Response to therapy may also be monitored with EBV-DNA. There is no standard therapy for PCNSL. Whole-brain radiation (WBRT) achieves CR in up to 50% but this is not translated to

increased survival, with median survival no more than 3 months. Deaths are generally related to opportunistic infections due to overwhelming immunosuppression at time of diagnosis. Even though many patients are unable to tolerate the full dose of radiation, the strongest predictors of outcome are performance status and the ability to deliver higher effective radiation doses.

A promising alternative to WBRT was studied in 15 patients using single-agent MTX intravenously at 3g/m². The mean CD4 count in these patients was 30/uL. Almost 50% had achieved CR with a median survival of 19 months and a relapse rate of only 14%⁶⁵. There is a survival benefit associated with the use of cART after diagnosis⁶⁶, and there is evidence that cART may increase the radio-sensitivity of B cells within the lymphoma^{67,68}. Given the very limited benefit of current modalities, patients should be referred to clinical trials.

Since there is universal association of EBV in HIV-associated PCNSL, therapeutic options which target the virus have been explored. In this regard it should be noted that EBV-specific allogeneic CTL have been shown to cross the blood brain barrier and induce tumour lysis. In the absence of an available study, either first-line WBRT or alternatively high-dose MTX with the option of WBRT consolidation should be considered. Concomitant HAART therapy to enhance the immune system is critical to successful outcomes

Classical Hodgkin lymphoma: HL is the most common type of non-AIDS defining tumor. The risk of developing HL in HIV patients is up to 11-18 times above the general population⁶⁹. It is associated with advanced disease and is more common in the intravenous drug group than in homosexual men. Its hallmark includes aggressive

clinical presentation with systemic B symptoms, widespread non-contiguous extranodal lesions and frequent bone marrow involvement (in up to 50% of cases). The morphological patterns are similar to those seen in patients without HIV infection, although with a greater proportion of the subtypes (mixed cellularity, lymphocyte depleted) with less favourable prognosis compared to the general population⁷⁰. As noted above, the greater proportion of mixed cellularity and lymphocyte depleted subtypes appears specifically related to severe immunocompromise in HIV, while HIV-infected patients with modest immunocompromise are more at risk for the development of the nodular sclerosis subtype.

The composition of the reactive inflammatory infiltrate in HIV-associated HL is often characterised by a predominance of CD8-positive T lymphocytes over CD4-positive lymphocytes, by contrast with the background in HL without HIV infection⁷⁰. This finding may simply reflect the depleted peripheral CD4 counts in this patient population. The cytological and phenotypic features of the Hodgkin Reed–Sternberg (HRS) cells in HIV-associated HL are similar to those in non-HIV associated HL. It has been determined that RS cells of all histologic categories of HIV-HD consistently display the BCL-6(-)/syn-1(+) phenotype and thus reflect post-GC B cells⁷¹.

The HRS cells typically express CD15 and CD30, express CD20 in a minor subset, and lack expression of CD45⁷⁰. In the vast majority of HIV associated HL there is coincident EBV infection. The latent EBV proteins EBNA-1, LMP-1, and LMP2A are expressed in the RS cells, the malignant cell population of this tumor⁷². RS cells are derived from B cells that have passed through the germinal center, as shown by the presence of somatic mutations in the rearranged Ig variable region of their immunoglobulin genes⁷³. LMP2A interferes with normal B-cell development, allows BCR-negative B cells to leave the bone marrow/colonize peripheral lymphoid organs⁷⁴, and induces a transcriptome pattern in B cells, which resembles that of HL RS cells⁷⁵. Following EBV infection, LMP2A is essential for the survival and continued proliferation of germinal center B cells lacking a functional B-cell receptor^{76,77}. LMP2A may therefore promote the survival of “crippled” germinal center B cells and could thus aid their development into RS cells (**Table 2, Table 3**).

LMP-1 may also induce an “HL-like” transcriptional program in germinal center B cells⁷⁸. Among the cellular genes up-regulated by LMP-1 in

HL cells is *bmi-1*, a polycomb family member known to cause lymphoma in transgenic mice and to down-regulate the ATM tumor suppressor⁷⁹. EBNA-1 was shown to induce CCL-20 secretion in RS cell lines and to thereby promote the migration of regulatory T cells, which could be envisaged to downmodulate EBV-specific T-cell responses⁸⁰.

This association with EBV is considerably stronger than that seen in HL in the non-HIV infected population. HIV-associated HL most often presents at an advanced clinical stage, with B symptoms, frequent extranodal disease, as bone marrow localization, and an aggressive course⁸¹. Unusual extranodal sites, such as the skin, lung and gastrointestinal tract may be involved⁸². These sites are essentially never involved by HL that is not associated with HIV.

HIV-HL patients have reduced CR rates and survival compared with the HIV negative population. In the early years post-HAART therapy the incidence of HIV-HL appeared to be in decline however two studies showed that the incidence may actually have increased^{83,84}.

The post-HAART era was also associated with an improvement in survival which was attributed to virological response to antiretroviral therapy and a reduction in HIV-associated mortality⁸⁵. In another study of 47 patients in the post-HAART era, the median survival was not reached compared with 19 months in the pre-HAART era^{86,87}.

Optimal therapy for HIV-HL has not been defined. Treatment regimes used are similar to those used in HL in the seronegative population^{88,89}.

Primary effusion lymphoma (PEL): PEL is a distinct clinicopathological entity occurring almost exclusively in HIV-infected patients. This lymphoma subtype comprises less than 5% of all HIV-associated NHL. Cases of this type were first described by Knowles et al in 1989⁹⁰, but its distinctive features were not fully recognised until after the identification of the Kaposi sarcoma-associated herpesvirus/human herpesvirus 8 (KSHV/HHV8) in 1994^{91,92}. PEL is a distinct type of B-cell non-Hodgkin lymphoma (NHL) that presents most frequently in body cavities as lymphomatous effusions without an associated tumor mass.

The tumor cells have large round to irregular nuclei with prominent nucleoli, and abundant deeply basophilic and occasionally vacuolated cytoplasm. These are described as immunoblastic/plasmablastic or anaplastic morphological features. Recent studies have

broadened the scope of PEL to include those presenting as a solid tumour mass with or without an associated effusion^{93,94,95}. The so-called “extracavitary” or “solid variant” of PEL most commonly involves the gastrointestinal tract or soft tissue, but can also involve lymph nodes. Some studies have suggested that the extracavitary variant of PEL has a slightly better prognosis when compared with cases presenting with effusion^{95,96}.

A defining property of PEL is its consistent association with KSHV infection. Most cases are also co-infected by EBV. It is believed that KSHV, rather than EBV, is a driving force in these tumors, as in PEL, at least 5 KSHV viral genes are expressed, which provide proliferative and antiapoptotic signals. In contrast, EBV has a restricted latency pattern of gene expression in PEL, where only EBNA1 and EBERs are expressed⁹⁷. However, the viral oncoprotein LMP-1 is generally not expressed^{94,95,96,97} (**Table 2, Table 3**).

The immunophenotypic features of PEL often make it difficult to confirm B-cell lineage, as the neoplasm usually lacks expression of most B-cell associated antigens including CD19, CD20, CD79a and immunoglobulins. The most frequently expressed antigens include those associated with activation or plasmacytic differentiation, such as CD30, CD45, EMA, CD71, MUM1, and CD138. Aberrant expression of Tcell associated antigens CD3 and CD7 has been reported^{98,99,100,101,102}.

B-cell origin of PELs, can be demonstrated by the presence of clonal immunoglobulin gene rearrangements. Evidence points toward a post-germinal center B-cell derivation, as most PELs contain somatic hypermutation of Ig genes as well as frequent somatic hypermutation of the noncoding region of the *BCL6* gene^{103,104}. Consistent with this notion is the expression of plasma cell markers such as CD138/Syndecan-1. Recently, gene expression analysis of PEL showed features most similar to AIDS immunoblastic lymphoma and multiple myeloma, again indicating a pre-plasma cell or “plasmablastic” profile¹⁰⁵.

Again, the exact role of EBV has been debated; but the fact that both viruses are detected together in most of the cases suggests that EBV may act as a cofactor in the initiating events (because it can immortalize and transform B cells in vitro and HHV-8 cannot), whereas HHV-8 may be the driving force for the tumor¹⁰⁶. With or without therapy, PEL is invariably associated with an adverse prognosis. There is limited data on the treatment of PEL. The destruction of local tissue

despite aggressive therapy leads to shortened survival¹⁰⁷.

Chemotherapy and radiotherapy may result in responses but these are seldom durable and survival is generally less than 12 months although a small series suggests the addition of high-dose MTX may improve outcome¹⁰⁷. Interestingly, a patient treated with a combination of zidovudine and α -interferon (α -IFN) entered into durable remission after only 5 days¹⁰⁸. Study of the primary tumour cells derived from this patient demonstrated that azidothymidine (AZT) blocked nuclear translocation of NF κ B and potentiated the pro-apoptotic effect of α -IFN (which induces another death receptor ligand, TRAIL). Further clinical studies of this combination are under way. In a murine system sirolimus showed promising activity which was in part mediated by inhibition of IL-10 signaling¹⁰⁸. Given the relative rarity of this lymphoma, patients should be enrolled in clinical trials where possible. Concomitant administration of HAART is advised and there are several reports of remission of PEL with use of HAART alone.

Plasmablastic lymphoma of the oral cavity type:

Plasmablastic lymphoma is a distinct type of diffuse large B-cell lymphoma that occurs most often in the oral cavity or jaw of HIV-infected individuals¹¹⁰. This rare lymphoma subtype accounts for 2.6% of HIV-related NHL¹¹¹. The first description designated this tumour as a lymphoma of the oral cavity⁶⁸; however, subsequent reports have described less frequent involvement of extraoral sites such as the anal cavity, gastrointestinal tract, lung, paranasal sinus, skin, spermatic cord, testicle, bone and lymph nodes¹¹²⁻¹¹⁸.

Regardless of the site of occurrence, plasmablastic lymphoma shows similar morphological and phenotypic features. The neoplastic cells are intermediate to large in size, with round nuclear contours and occasional multinucleation. Plasmacytic differentiation is usually apparent, with a cytological spectrum including a minor population of small plasmacytoid cells with condensed chromatin ranging to large cells with dispersed chromatin, prominent central nucleoli and abundant basophilic cytoplasm with a paranuclear hof^{112,113,114}. The neoplastic population generally expresses CD45 and plasmacytic markers such as CD138, EMA and MUM1, and usually lacks expression of pan-B-cell antigens such as CD20 and PAX5.1^{110,116}. In early reports, slightly more than 50% of cases were EBER positive as shown by in situ hybridisation studies¹¹⁰ in more

recent series all cases of plasmablastic lymphoma have been shown to be EBER positive¹¹⁵, 73 EBER-positive cases generally lack expression of EBNA2 and LMP-1^{115,116} (**Table 2, Table 3**). HHV8 infection is not implicated in the pathogenesis of plasmablastic lymphoma, with all cases negative for LNA1 when tested by immunohistochemistry. While there is morphological and phenotypic overlap with anaplastic myeloma, extramedullary presentation and frequent EBV infection are distinctive features. A potential role for EBV in the pathogenesis of the disease remains unknown, especially with the highly restricted latency expression pattern. Despite the use of aggressive chemotherapy and HAART the prognosis remains poor¹¹⁹.

Polymorphic B-cell Lymphoma (PTLD-like):

HIV infection results in a reduction of T-cell immunity similar to that iatrogenically induced in transplant patients. Is not surprising that polymorphic lymphoid proliferations resembling post-transplant lymphoproliferative disorders (PTLD) have been reported in HIV-infected adults and children. According to the WHO classification, they are divided into early lesions (reactive plasmacytic hyperplasia and mononucleosis-like syndrome), polymorphic lesions, monomorphic lesions, and Hodgkin-like lesions¹²⁰. Similarly to PTLD, these infiltrates are often associated with EBV infection. By contrast with HIV-associated lymphoma, these polymorphic infiltrates often show more limited disease distribution, lack oncogene and tumour suppressor gene alterations, and may be polyclonal or show a minor B-cell clone in a polyclonal background. Regression of polymorphic B-cell lymphoma in an HIV-infected patient after anti-retroviral therapy has been reported¹²¹.

EBV has been linked to most PTLDS, with a near 100% association in the early-occurring cases (within a year) and in PTLD-associated Hodgkin lymphoma¹²². The EBV-negative PTLDS constitute approximately 20% of all cases, have a tendency to late occurrence and have an unknown etiology. Type III latency is exhibited by the EBV-positive B cells in PTLD, although some studies have reported a more restricted latency pattern¹²³. The wide expression of the latent EBV-encoded proteins strongly suggests an important role that EBV may play in the oncogenic process (**Table 2, Table 3**).

The mechanism by which EBV is thought to contribute to the pathogenesis of PTLD is similar to its presumed role in Hodgkin lymphoma. Because approximately 50% of PTLD cases are derived from

GC B cells lacking a functional BCR because of certain crippling mutations, and because these cells manage to escape apoptosis despite lacking antigen affinity, it is believed that EBV aids in rescuing these cells from an imminent programmed cell death^{124,125}. As in Hodgkin cases, LMP1 and LMP2A may replace survival signals induced by activated BCR and CD40 receptors and also activate the NF- κ B signaling pathway, inducing proliferation of neoplastic cells. The decreased cytotoxic T-cell surveillance because of immunosuppression in PTLD patients is also believed to greatly facilitate the actions of EBV. The similar role that EBV is thought to play in inducing the survival and neoplastic transformation of infected GC cells in both PTLD and Hodgkin lymphoma, in addition to the near 100% EBV positivity in PTLD-associated Hodgkin lymphoma, has led some investigators to speculate a connection between the 2 diseases and the possibility that EBV infection and its GC effects may be the initiating role in the pathogenesis of both entities¹²⁴.

Conclusions: HIV-associated lymphomas represent a particular setting characterizing specific pathogenetic models prevalently driven by EBV and by immunodeficiency. The impact of combined antiretroviral therapy has substantially changed the risk and prognosis of lymphoma in HIV-infected population, as well as the relationship with the natural history of HIV disease. As a consequence of cART, many authors now strongly recommend that patients with lymphoma and HIV infection should be treated as patients with lymphoma of the general population.

In fact, due to the improvement of morbidity and mortality related with cart exposure, more aggressive treatment protocols can be taken into consideration, on the bases of the results in terms of efficacy and tolerability reported in the general population, such as the use of high-dose chemotherapy in combination with PBSC transplantation in HIV-NHL which showed response rates similar to those obtained in HIV-negative patients.

The concurrent use of antineoplastic chemotherapy and cART should be considered a potential advantage for tumor prognosis and for reducing risk of toxicities associated to antineoplastic drugs, even though concerns due to drug-drug interactions could be suggested. In perspectives, the molecular and epidemiological linkage between AIDS-related malignancies and EBV-infection suggests that viral gene products would be potential targets for

molecular-targeted chemotherapy. Detailed understanding of the EBV lifecycle and related cancers at the molecular level may lead to the development of novel strategies of molecular-

targeted cancer chemotherapy to specific viral oncogenes to which the lymphoma cells are addicted, and that will provide therapeutic benefits.

References

- Ledergerber B, Telenti A, Egger M. Risk of HIV related Kaposi's sarcoma and Non-Hodgkin's lymphoma with potent antiretroviral therapy: prospective cohort study. *Br Med J* 1999;319:23-24.
- Stebbing J, Gazzard B, Mandalia S, et al. Antiretroviral treatment regimens and immune parameters in the prevention of systemic AIDS-related non-Hodgkin's lymphoma. *J Clin Oncol* 2004;22:2177-2183.
- May T, Lewden C, Bonnet F, et al. Causes and characteristics of death among HIV-1 infected patients with immunovirologic response to antiretroviral treatment. *Presse Med* 2004;33:1487-1492.
- Besson C, Goubar A, Gabarre J, et al. Changes in AIDS-related lymphoma since the era of highly active antiretroviral therapy. *Blood* 2001;98:2339-2344.
- Clifford GM, Polesel J, Rickenbach M, et al. Cancer risk in the Swiss HIV cohort study: associations with immunodeficiency, smoking, and highly active antiretroviral therapy. *J Natl Cancer Inst* 2005;97:425-432.
- Engels EA, Pfeiffer RM, Goedert JJ, et al. Trends in cancer risk among people with AIDS in the United States 1980-2002. *AIDS* 2006;20:1645-1654.
- Herndier B, Shiramizu B, Jewett N, et al. Acquired immunodeficiency syndrome associated T-cell lymphoma: evidence for human immunodeficiency virus type 1-associated T-cell transformation. *Blood* 1992;79:1768-74.
- Bellan C, Lazzi S, DeFalco G, et al. Burkitt's lymphoma: new insights into molecular pathogenesis. *J Clin Pathol* 2003;56:188-93.
- Tirelli U, Errante D, Dolcetti R, Gloghini A, Serraino D, Vaccher E, Franceschi S, Boiocchi M, Carbone A. Hodgkin disease and human immunodeficiency virus infection: clinicopathologic and virologic features of 114 patients from the Italian Cooperative Group on AIDS and Tumors. *J. Clin. Oncol.* **13**, 1758-1767 (1995).
- INSERM U720: Epidemiologie Clinique et Traitement de l'Infection a' VIH: Retour d'Informations Clinico-Epidemiologiques. Septembre 2007, <http://www.code.fr>
- Tran H, Nourse L, Hall S, Green M, et al. *Blood Reviews* 2008;22:261-281.
- Raphael M, Borisch B, Jaffe E. Lymphomas associated with infection by the human immunodeficiency virus (HIV). In: Jaffe E, Harris N, Stein H, Vardiman J, eds. World Health Organization classification of tumours, pathology and genetics of tumours of haematopoietic and lymphoid tissues. Lyon, France: IARC Press, 2001:260-3.
- Jarrett R. Viruses and lymphoma/leukaemia. *J Pathol* 2006;208:176-86.
- Knowles D. Etiology and pathogenesis of AIDS-related non-Hodgkin's lymphoma. *Hematol Oncol Clin N Am* 2003;17:785-820.
- Laurence J, Astrin S. Human immunodeficiency virus induction of malignant transformation in human B lymphocytes *Proc Natl Acad Sci* 1991;88:7635-9
- Kersten, M. J., Klein, M. R., Holwerda, A. M., Miedema, F. & van Oers, M. H. Epstein-Barr virus-specific cytotoxic T cell responses in HIV-1 infection: different kinetics in patients progressing to opportunistic infection or non-Hodgkin's lymphoma. *J. Clin. Invest.* **99**, 1525-1533 (1997).
- Van Baarle, D. et al. Dysfunctional Epstein-Barr virus (EBV)-specific CD8+ T lymphocytes and increased EBV load in HIV-1 infected individuals progressing to AIDS-related non-Hodgkin lymphoma. *Blood* **98**, 146-155 (2001).
- Van Baarle, D. et al. Lack of Epstein-Barr virus- and HIV-specific CD27-CD8+ T cells is associated with progression to viral disease in HIV-infection. *AIDS* **16**, 2001-2011
- Mbulaiteye, S. M., Biggar, R. J., Goedert, J. J. & Engels, E. A. Immune deficiency and risk for malignancy among persons with AIDS. *J. Acquir. Immune. Defic. Syndr.* **32**, 527-533 (2003).
- Muñoz A, Schragger LK, Bacellar H, Speizer I, Vermund SH, Detels R, Saah AJ, Kingsley LA, Seminara D, Phair JP. Trends in the incidence of outcomes defining acquired immunodeficiency syndrome (AIDS) in the Multicenter AIDS Cohort Study: 1985-1991. *Am. J. Epidemiol.* **137**, 1985-1991 (1993).
- Kirk O, Pedersen C, Cozzi-Lepri A, Antunes F, Miller V, Gatell JM, Katlama C, Lazzarin A, Skinhøj P, Barton SE; EuroSIDA Study Group. Non-Hodgkin lymphoma in HIV-infected patients in the era of highly active antiretroviral therapy. *Blood* **98**, 3406-3412 (2001).
- Grossman, Z., Meier-Schellersheim, M., Sousa, A. E., Victorino, R. M. & Paul, W. E. CD4+ T-cell depletion in HIV infection: are we closer to understanding the cause? *Nature Med.* **8**, 319-323 (2002).
- Hellerstein, M. K. et al. Subpopulations of long-lived and short-lived T cells in advanced HIV-1 infection. *J. Clin. Invest.* **112**, 956-966 (2003).
- Kovacs JA, Lempicki RA, Sidorov IA, Adelsberger JW, Herpin B, Metcalf JA, Sereti I, Polis MA, Davey RT, Tavel J, Falloon J, Stevens R, Lambert L, Dewar R, Schwartzentruber DJ, Anver MR, Baseler MW, Masur H, Dimitrov DS, Lane HC. Identification of dynamically distinct subpopulations of T lymphocytes that are differentially affected by HIV. *J. Exp. Med.* **194**, 1731-1741 (2001).
- Ribeiro, R. M., Mohri, H., Ho, D. D. & Perelson, A. S. *In vivo* dynamics of T cell activation, proliferation, and death in HIV-1 infection: why are CD4+ but not CD8+ T cells depleted? *Proc. Natl Acad. Sci. USA* **99**, 15572-15577 (2002)
- Pellici PG, Knowles DM 2nd, Arlin ZA, Wieczorek R, Luciw P, Dina D, Basilico C, Dalla-Favera R. Multiple monoclonal B cell expansions and *c-myc* oncogene rearrangements in acquired immune deficiency syndrome-related lymphoproliferative disorders. Implications for lymphomagenesis. *J. Exp. Med.* **164**, 2049-2060 (2003).
- Carbone, A. Emerging pathways in the development of AIDS-related lymphomas. *Lancet Oncol.* **4**, 22-29 (2003).
- Rezk SA, Weiss LM. Epstein-Barr virus-associated lymphoproliferative disorders. *Hum Pathol* 2007;38:1293-1304.
- Said JW. Immunodeficiency-related Hodgkin lymphoma and its mimics. *Adv Anat Pathol.* 2007;14:189-194.
- Carbone A, Gloghini A, Larocca LM, et al. Human immunodeficiency virus-associated Hodgkin's disease derives from post-germinal center B cells. *Blood.* 1999;93:2319-2326.
- Spina M, Tirelli U, Zagonel V, et al. Burkitt's lymphoma in adults with and without human immunodeficiency virus infection. *Cancer* 1998;82:766-74.
- Gold J, Castella A, Zalusky R. B-cell acute lymphocytic leukemia in HIV-antibodypositive patients. *Am J Hematol* 1989;32:200-4.
- Young LS, Rickinson AB. Epstein-Barr virus: 40 years on. *Nat Rev Cancer.* 2004;4:757-768.

34. Kamranvar SA, Gruhne B, Szeles A, Masucci MG. Epstein-Barr virus promotes genomic instability in Burkitt's lymphoma. *Oncogene*. 2007;26: 5115-5123.
35. Grogg KL, Miller RF, and Dogan A. HIV infection and lymphoma: *J. Clin. Pathol*. 2007;60:1365-1372.
36. Bellan C, Lazzi S, DeFalco G, et al. Burkitt's lymphoma: new insights into molecular pathogenesis. *J Clin Pathol* 2003;56:188-93
37. Hamilton-Dutoit SJ, Rea D, Raphael M, et al. Epstein-Barr virus-latent gene expression and tumor cell phenotype in acquired immunodeficiency syndrome-related non-Hodgkin's lymphoma: correlation of lymphoma phenotype with three distinct patterns of viral latency. *Am J Pathol*. 1993;143:1072-1085.
38. Carbone A, Tirelli U, Glohini A, Volpe R, Boiocchi M. Human immunodeficiency virus-associated systemic lymphomas may be subdivided into two main groups according to Epstein-Barr viral latent gene expression. *J Clin Oncol*. 1993;11:1674-1681.
39. Young LS, Rickinson AB. Epstein-Barr virus: 40 years on. *Nat Rev Cancer*. 2004;4:757-768.
40. Wang D, Liebowitz D, Kieff E. An EBV membrane protein expressed in immortalized lymphocytestransforms established rodent cells. *Cell*. 1985;43:831-840.
41. Kulwichit W, Edwards RH, Davenport EM, Baskar JF, Godfrey V, Raab-Traub N. Expression of the Epstein-Barr virus latent membrane protein 1 induces B cell lymphoma in transgenic mice. *Proc Natl Acad Sci U S A*. 1998;95:11963-11968.
42. Dirmeier U, Neuhierl B, Kilger E, Reischbach G, Sandberg ML, Hammerschmidt W. Latent membrane protein 1 is critical for efficient growth transformation of human B cells by Epstein-Barr virus. *Cancer Res*. 2003;63:2982-2989.
43. Huen DS, Henderson SA, Croom-Carter D, Rowe M. The Epstein-Barr virus latent membrane protein- 1 (LMP1) mediates activation of NF-kappa B and cell surface phenotype via two effector regions in its carboxy-terminal cytoplasmic domain. *Oncogene*. 1995;10:549-560.
44. Devergne O, Hummel M, Koeppen H, et al. A novel interleukin-12 p40-related protein induced by latent Epstein-Barr virus infection in B lymphocytes. *J Virol*. 1996;70:1143-1153.
45. Izumi KM, Kaye KM, Kieff ED. The Epstein-Barr virus LMP1 amino acid sequence that engages tumor necrosis factor receptor associated factors is critical for primary B lymphocyte growth transformation. *Proc Natl Acad Sci U S A*. 1997;94: 1447-1452.
46. Brinkmann MM, Schulz TF. Regulation of intracellular signalling by the terminal membrane proteins of members of the Gammaherpesvirinae. *J Gen Virol*. 2006;87:1047-1074.
47. Gaidano G, Carbone A, Dalla-Favera R. Pathogenesis of AIDS-related lymphomas: molecular and histogenetic heterogeneity. *Am J Pathol*. 1998;152:623-630.
48. Guasparri I, Bubman D, Cesarman E. EBV LMP2A affects LMP1-mediated NF-kappaB signaling and survival of lymphoma cells by regulating TRAF2 expression. *Blood*. 2008;111:3813-3820
49. Rowe M, Young LS, Crocker J, Stokes H, Henderson S, Rickinson AB. Epstein-Barr virus(EBV)-associated lymphoproliferative disease in the SCID mouse model: implications for the pathogenesis of EBV-positive lymphomas in man. *J Exp Med*. 1991;173:147-158.
50. Flinn IW, Ambinder RF. AIDS primary central nervous system lymphoma. *Curr Opin Oncol* 1996;8:373-6.
51. Besson C, Goubar A, Gabarre J et al. Changes in AIDS-related lymphoma since the era of highly active antiretroviral therapy. *Blood* 2001;98:2339-4
52. Raez LE, Patel P, Feun L, Restrepo A, Raub Jr WA, Cassileth PA. Natural history and prognostic factors for survival in patients with acquired immune deficiency syndrome (AIDS)-related primary central nervous system lymphoma (PCNSL). *Crit Rev Oncog* 1998;9:199-208
53. Cohen JL. Clinical aspects of Epstein-Barr virus infection. In: Robertson ES, editor. *Epstein-Barr virus*. Norfolk: Caister Academic press; 2005. p. 35-55.
54. Cinque P, Vago L, Dahl H, et al. Polymerase chain reaction on cerebrospinal fluid for diagnosis of virus-associated opportunistic diseases of the central nervous system in HIV-infected patients. *AIDS* 1996;10:951-8.
55. Cingolani A, De Luca A, Larocca LM, et al. Minimally invasive diagnosis of acquired immunodeficiency syndrome-related primary central nervous system lymphoma. *J Natl Cancer Inst* 1998;90:364-369.
56. Ivers LC, Kim AY, Sax PE. Predictive value of polymerase chain reaction of cerebrospinal fluid for detection of Epstein-Barr virus to establish the diagnosis of HIV-related primary central nervous system lymphoma. *Clin Infect Dis* 2004;38:1629-32.
57. MacMahon EM, Glass JD, Hayward SD, et al. Epstein-Barr virus in AIDS-related primary central nervous system lymphoma. *Lancet* 1991;338:969-973.
58. Larocca LM, Capello D, Rinelli A, et al. The molecular and phenotypic profile of primary central nervous system lymphoma identifies distinct categories of the disease and is consistent with histogenetic derivation from germinal center-related B cells. *Blood* 1998;92:1011-1019.
59. Maher EA, Fine HA. Primary CNS lymphoma. *Semin Oncol* 1999;26:346-56.
60. Johnson BA, Fram EK, Johnson PC, Jacobowitz R. The variable MR appearance of primary lymphoma of the central nervous system: comparison with histopathologic features. *AJNR Am J Neuroradiol* 1997;18:563-72.
61. Antinori A, Ammassari A, De Luca A, et al. Diagnosis of AIDS-related focal brain lesions: a decision-making analysis based on clinical and neuroradiologic characteristics combined with polymerase chain reaction assays in CSF. *Neurology* 1997;48:687-694.
62. Antinori A, De Rossi G, Ammassari A et al. Value of combined approach with thallium-201 single-photon emission computed tomography and Epstein-Barr virus DNA polymerase chain reaction in CSF for the diagnosis of AIDS-related primary CNS lymphoma. *J Clin Oncol* 1999;17: 554-60.
63. Castagna A, Cinque P, d'Amico A, Messa C, Fazio F, Lazzarin A. Evaluation of contrast-enhancing brain lesions in AIDS patients by means of Epstein-Barr virus detection in cerebrospinal fluid and ²⁰¹thallium single photon emission tomography. *AIDS* 1997;11:1522-3.
64. Antinori A, Ammassari A, Luzzati R, et al. Role of brain biopsy in the management of focal brain lesions in HIV-infected patients. Gruppo Italiano Cooperativo AIDS & Tumori. *Neurology* 2000;54:993-997.
65. Jacomet C, Girard PM, Lebrette MG, Farese VL, Monfort L, Rozenbaum W. Intravenous methotrexate for primary central nervous system non-Hodgkin's lymphoma in AIDS.
66. Newell ME, Hoy JF, Cooper SG et al. Human immunodeficiency virus-related primary central nervous system lymphoma: factors influencing survival in 111 patients. *Cancer* 2004;100:2627-36.
67. Hoffmann C, Tabrizian S, Wolf E et al. Survival of AIDS patients with primary central nervous system lymphoma is dramatically improved by HAART-induced immune recovery. *AIDS* 2001;15:2119-27.
68. Pajonk F, McBride WH. Survival of AIDS patients with primary central nervous system lymphoma may be improved by the radiosensitizing effects of highly active antiretroviral therapy. *AIDS* 2002;16:1195-6.
69. Goedert JJ, Cote TR, Virgo P et al. Spectrum of AIDS-associated malignant disorders. *Lancet*. 1998;351: 1833-9.
70. Thompson L, Fisher M, Chu W, et al. HIV-associated Hodgkin lymphoma. *Am J Clin Pathol* 2004;121:727-38.
71. Carbone A, Glohini A, Larocca LM, et al. Human immunodeficiency virus-associated Hodgkin's disease derives from post-germinal center B cells. *Blood* 1999;93:2319-2326.
72. Young LS, Murray PG. Epstein-Barr virus and oncogenesis: from latent genes to tumours. *Oncogene*. 2003;22:5108-5121.
73. Kuppers R, Rajewsky K, Zhao M, et al. Hodgkin disease: Hodgkin and Reed-Sternberg cells picked from histological sections show clonal immunoglobulin gene rearrangements

- and appear to be derived from B cells at various stages of development. *Proc Natl Acad Sci U S A.* 1994;91:10962-10966.
74. Caldwell RG, Wilson JB, Anderson SJ, Longnecker R. Epstein-Barr virus LMP2A drives B cell development and survival in the absence of normal B cell receptor signals. *Immunity.* 1998;9:405-411.
 75. Portis T, Dyck P, Longnecker R. Epstein-Barr Virus (EBV) LMP2A induces alterations in gene transcription similar to those observed in Reed-Sternberg cells of Hodgkin lymphoma. *Blood* 2003;102:4166-4178
 76. Mancao C, Altmann M, Jungnickel B, Hammerschmidt W. Rescue of "crippled" germinal center B cells from apoptosis by Epstein-Barr virus. *Blood.* 2005;106:4339-4344.
 77. Mancao C, Hammerschmidt W. Epstein-Barr virus latent membrane protein 2A is a B-cell receptor mimic and essential for B-cell survival. *Blood.* 2007;110:3715-3721.
 78. Vockerodt M, Morgan S, Kuo M, et al. The Epstein-Barr virus oncoprotein, latent membrane protein-1, reprograms germinal center B cells towards a Hodgkin's Reed-Sternberg-like phenotype. *J Pathol.* 2008;216:83-92.
 79. Dutton A, Woodman CB, Chukwuma MB, et al. Bmi-1 is induced by the Epstein-Barr virus oncogene LMP1 and regulates the expression of viral target genes in Hodgkin lymphoma cells. *Blood* 2007;109:2597-2603.
 80. Baumforth KR, Birgersdotter A, Reynolds GM, et al. Expression of the Epstein-Barr virus-encoded Epstein-Barr virus nuclear antigen 1 in Hodgkin's lymphoma cells mediates up-regulation of CCL20 and the migration of regulatory T cells. *Am J Pathol.* 2008;173:195-204.
 81. Thompson L, Fisher M, Chu W, et al. HIV-associated Hodgkin lymphoma. *Am J Clin Pathol* 2004;121:727-38.
 82. Doweiko J, Dezube B, Pantanowitz L. Unusual sites of Hodgkin's lymphoma. *J Clin Oncol* 2004;22:4227-31
 83. Biggar RJ, Jaffe ES, Goedert JJ, Chaturvedi A, Pfeiffer R, Engels EA. Hodgkin lymphoma and immunodeficiency in persons with HIV/AIDS. *Blood* 2006;108:3786-91
 84. Herida M, Mary-Krause M, Kaphan R et al. Incidence of non-AIDS-defining cancers before and during the highly active antiretroviral therapy era in a cohort of human immunodeficiency virus-infected patients. *J Clin Oncol* 2003;21:3447-53.
 85. Palella Jr FJ, Delaney KM, Moorman AC et al. Declining morbidity and mortality among patients with advanced human immunodeficiency virus infection. HIV Outpatient Study Investigators. *N Engl J Med.* 1998;338:853-60.
 86. Gerard L, Galicier L, Boulanger E et al. Improved survival in HIV-related Hodgkin's lymphoma since the introduction of highly active antiretroviral therapy. *Aids* 2003;17:81-7.
 87. Glaser SL, Clarke CA, Gullely ML et al. Population-based patterns of human immunodeficiency virus-related Hodgkin lymphoma in the Greater San Francisco Bay Area, 1988-1998. *Cancer* 2003;98:300-9.
 88. Hartmann P, Rehwald U, Salzberger B et al. BEACOPP therapeutic regimen for patients with Hodgkin's disease and HIV infection. *Ann Oncol* 2003;14:1562-9.
 89. Spina M, Gabarre J, Rossi G et al. Stanford V regimen and concomitant HAART in 59 patients with Hodgkin disease and HIV infection. *Blood* 2002;100:1984-8.
 90. Knowles D, Inghirami G, Ubriaco A, et al. Molecular genetic analysis of three AIDS-associated neoplasms of uncertain lineage demonstrates their B-cell derivation and the possible pathogenetic role of the Epstein-Barr virus. *Blood* 1989;73:792-9.
 91. Nador R, Cesarman E, Chadburn A, et al. Primary effusion lymphoma: a distinct clinicopathologic entity associated with the Kaposi's sarcoma-associated herpesvirus. *Blood* 1996;88:645-56.
 92. Cesarman E, Chang Y, Moore P, et al. Kaposi's sarcoma-associated herpesviruslike DNA sequences in AIDS-related body-cavity-based lymphomas. *N Engl J Med* 1995;332:1186-91.
 93. Chang Y, Cesarman E, Moore P, et al. Identification of herpesvirus-like DNA sequences in AIDS-associated Kaposi's sarcoma. *Science* 1994;266:1865-9.
 94. Carbone A, Gloghini A, Vaccher E, et al. Kaposi's sarcoma-associated herpesvirus/human herpesvirus type 8-positive solid lymphomas. *J Mol Diagn* 2005;7:17-27.
 95. Chadburn A, Hyjek E, Mathew S, et al. KSHV-positive solid lymphomas represent an extra-cavitary variant of primary effusion lymphoma. *Am J Surg Pathol* 2004;28:1401-16.
 96. Deloose S, Smit L, Pals F, et al. High incidence of Kaposi sarcoma-associated herpesvirus infection in HIV-related solid immunoblastic/plasmablastic diffuse large B-cell lymphoma. *Leukemia* 2005;19:851-5.
 97. Carbone A, Cesarman E, Spina M, Gloghini A and Thomas F. Schulz: HIV-associated lymphomas and gamma-herpesviruses: *Blood* ; 2009 113: 1213-1224
 98. Boulanger E, Hermine O, Ferman J-P, et al. Human herpesvirus 8 (HHV8)-associated peritoneal primary effusion lymphoma (PEL) in two HIV-negative elderly patients. *Am J Hematol* 2004;76:88-91.
 99. Oksenhendler E, Boulanger E, Galicier L, et al. High incidence of Kaposi sarcoma-associated herpesvirus-related non-Hodgkin lymphoma in patients with HIV infection and multicentric Castleman disease. *Blood* 2002;99:2331-6.
 100. Said J, Shintaku I, Asou H, et al. Herpesvirus 8 inclusions in primary effusion lymphoma: report of a unique case with T-cell phenotype. *Arch Pathol Lab Med* 2001;123:257-60.
 101. Beaty M, Kumar S, Sorbara L, et al. A biphenotypic human herpesvirus 8-associated primary bowel lymphoma. *Am J Surg Pathol* 1997;21:719-24
 102. Gaidano G, Capello D, Cilia AM, et al. Genetic characterization of HHV-8/KSHV-positive primary effusion lymphoma reveals frequent mutations of BCL6: implications for disease pathogenesis and histogenesis. *Genes Chromosomes Cancer.* 1999;24:16-23.
 103. Matolcsy A, Nador RG, Cesarman E, Knowles DM. Immunoglobulin VH gene mutational analysis suggests that primary effusion lymphomas derive from different stages of B cell maturation. *Am J Pathol.* 1998;153:1609-1614.
 104. Jenner RG, Maillard K, Cattini N. Kaposi's sarcoma-associated herpesvirus-infected primary effusion lymphoma has a plasma cell gene expression profile. *Proc Natl Acad Sci U S A.* 2003;100:10399-10404.
 105. Fan W, Bubman D, Chadburn A, Harrington Jr WJ, Cesarman E, Knowles DM. Distinct subsets of primary effusion lymphoma can be identified based on their cellular gene expression profile and viral association. *J Virol* 2005;79:1244-51.
 106. Simonelli C, Spina M, Cinelli R et al. Clinical features and outcome of primary effusion lymphoma in HIV-infected patients: a single-institution study. *J Clin Oncol* 2003;21:3948-54.
 107. Boulanger E, Daniel MT, Agbalika F, Oksenhendler E. Combined chemotherapy including high-dose methotrexate in KSHV/HHV8-associated primary effusion lymphoma. *Am J Hematol* 2003;73:143-8.
 108. Ghosh SK, Wood C, Boise LH et al. Potentiation of TRAIL-induced apoptosis in primary effusion lymphoma through azidothymidine-mediated inhibition of NF-kappa B. *Blood* 2003;101:2321-7.
 109. Sin SH, Roy D, Wang L et al. Rapamycin is efficacious against primary effusion lymphoma (PEL) cell lines in vivo by inhibiting autocrine signaling. *Blood* 2007;109:2165-73.
 110. Delecluse H, Anagnostopoulos I, Dallenbach F, et al. Plasmablastic lymphomas of the oral cavity: a new entity associated with the human immunodeficiency virus infection. *Blood* 1997;89:1413-20.
 111. Folk G, Abbondanzo S, Childers E, et al. Plasmablastic lymphoma: a clinicopathologic correlation. *Ann Diagn Pathol* 2006;10:8-12.
 112. Tavora F, Gonzalez-Cuyar L, Chen-Chih J, et al. Extra-oral plasmablastic lymphoma: report of a case and review of literature. *Human Pathol* 2006;37:1233-6.
 113. Schichman S, McClure R, Schaefer R, et al. HIV and plasmablastic lymphoma manifesting in sinus, testicles, and bones: a further expansion of the disease spectrum. *Am J Hematol* 2004;77:291-5.

114. Chetty R, Hlatswayo N, Muc R, et al. Plasmablastic lymphoma in HIV+ patients: an expanding spectrum. *Histopathology* 2003;42:605–9.
115. Dong H, Scadden D, de Leval L, et al. Plasmablastic lymphoma in HIV-positive patients: an aggressive Epstein-Barr virus-associated extramedullary plasmacytic neoplasm. *Am J Surg Pathol* 2005;29:1633–41.
116. Lin Y, Rodrigues G, Turner J, et al. Plasmablastic lymphoma of the lung: report of a unique case and review of the literature. *Arch Pathol Lab Med* 2001;125:282–5.
117. Pruneri G, Graziadei G, Ermellino L, et al. Plasmablastic lymphoma of the stomach. *Haematologica* 1998;83:87–9.
118. Hausermann P, Khanna N, Buess M, et al. Cutaneous plasmablastic lymphoma in an HIV-positive male: an unrecognized cutaneous manifestation. *Dermatology* 2004;208:287–90.
119. Castillo L, Pantanowitz L, Dezube BJ. HIV-associated plasmablastic lymphoma: lessons learned from 112 published cases. *Am J Hematol*. 2008 Oct;83(10):763-4.
120. Harris NL, Swerdlow SH, Frizzera G, Knowles DM. Post transplant lymphoproliferative disorders. In: Jaffe ES, Harris NL, Stein H, Vardiman JW, editors. *World Health Organization classification of tumors. Pathology and genetics of tumors of hematopoietic and lymphoid tissues*. Lyon: IARC Press; 2001. p. 264-70.
121. Martin S, Zukerberg L, Robbins G. Reactive Epstein-Barr virus-related polyclonal lymphoproliferative disorder in a patient with AIDS. *Clin Infect Dis* 2005;41:e76–9.
122. Thompson MP, Kurzrock R. Epstein-Barr virus and cancer. *Clin Cancer Res* 2004;10:803-21 .
123. Brink AA, Dukers DF, van den Brule AJ, et al. Presence of Epstein- Barr virus latency type III at the single cell level in posttransplantation lymphoproliferative disorders and AIDS related lymphomas. *J Clin Pathol* 1997;50:911-8.
124. Timms JM, Bell A, Flavell JR, et al. Target cells of Epstein-Barr-virus (EBV)-positive post-transplant lymphoproliferative disease: similarities to EBV-positive Hodgkin's lymphoma. *Lancet* 2003;361:217-23.
125. Capello D, Rossi D, Gaidano G. Post-transplant lymphoproliferative disorders: molecular basis of disease histogenesis and pathogenesis. *Hematol Oncol* 2005;23:61-7