

Scientific Letter

In vitro Activity of Cefiderocol Against Carbapenem-Resistant *Acinetobacter baumannii* Clinical Isolates: a Single Center Experience

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To the editor.

The introduction should briefly place the study in a broad context and highlight why it is Infections caused by multi-drug resistant (MDR) Gram-negative bacteria represent one of the main threats to human health worldwide.¹ Since the beginning of the SARS-CoV-2 pandemic, a significant increase of severe infections due to MDR ESKAPE bacteria was observed in our Institution, in particular, due to carbapenem-resistant Acinetobacter baumannii (CRAB).²⁻⁴ Cefiderocol (CFDC), approved in 2019 to treat infections sustained by aerobic Gram-negative bacteria, is a novel siderophore cephalosporin with broad-spectrum activity and clinical efficacy against CRAB, Pseudomonas aeruginosa and Stenotrophomonas maltophilia.5,6 However, since its introduction in clinical practice, CFDC-resistant Gram-negative bacterial isolates have been reported.7

Herein, we evaluated the *in vitro* activity of CFDC against CRAB bloodstream strains isolated in our Teaching Hospital during the last three years.

Materials and Methods. This study was conducted in the Microbiology Laboratory "*Mater Domini*" Teaching Hospital, Catanzaro, Italy. CRAB isolates were recovered from blood samples of patients hospitalized between 2020 and 2022 and diagnosed with CRAB bloodstream infections. Only the first CRAB strain isolated from each patient was included.

Bacterial isolation and identification and Antimicrobial Susceptibility Testing. CRAB isolates were identified using matrix-assisted laser-desorption ionization timeof-flight mass spectrometry (MALDI-TOF) and Vitek®2 System (bioMérieux, Italy). Antibiotic susceptibility tests for meropenem, amikacin, and trimethoprim/sulfamethoxazole were performed with the Vitek®2 System (whereas the determination of colistin resistance was obtained by broth microdilution according to EUCAST guidelines.8

The Kirby-Bauer disc diffusion test on regular unsupplemented Mueller-Hinton agar (Liofilchem S.R.L., Roseto degli Abruzzi, Italy) was used to assess sensitivity to CFDC, using discs impregnated with 30 micrograms of drug supplied by Liofilchem[®] (Liofilchem S.R.L., Roseto degli Abruzzi, Italy). Of note, EUCAST evaluated (August 2022) cefiderocol 30 µg disks and regular unsupplemented MH agars from Liofilchem, and no warning concerning their use was reported (unlike other products marketed by other manufacturers).⁹ Well-isolated colonies were suspended in saline from an overnight agar plate to achieve a 0.5 McFarland standard turbidity. The inoculum was streaked with a sterile cotton swab over the entire area of the Mueller-Hinton (MH) agar plate. After that, the disc was firmly applied to the surface of the inoculated agar plate and incubated at 35±1 °C for 16–20 h. The diameters of the disk areas were read using the innermost colony-free area. The results were interpreted using the European Committee on Antimicrobial Susceptibility Testing (EUCAST) and Clinical and Laboratory Standards Institute (CLSI) experimental breakpoints.9,10 The EUCAST defined the clinical breakpoints for CFDC against A. baumannii complex as susceptible in the range of ≥ 17 mm and resistant of < 17mm, while the CLSI ranges for the determination of susceptibility and resistance are ≥ 15 mm, and ≤ 15 mm, respectively.

Ethical Statement. The present retrospective study is based on clinical isolates stored in an anonymous archive without association with clinical data. For this reason, ethics and consent to participate are not applicable. The study was conducted using retrospectively collected and anonymized data according to the Declaration of Helsinki and principles of good clinical practice.

Table 1. Summary of cefiderocol disk diffusion susceptibility among analysed carbapenem-resistant Acinetobacter baumannii bacterial isolates.

		EUCAST brea zone diame	-	CLSI breakpoints disk zone diameter (mm)		
Carba	penem-resistant Acinetobacter	S	R	S	R	
	aumannii isolates (N=62)	≥17	<17	≥15	<15	
		% (1	N)	% (N)		
		72.5 (45)	27.5 (17)	83.9 (52)	16.1 (10)	

Abbreviations: CLSI, Clinical and Laboratory Standards Institute; EUCAST, European Committee on Antimicrobial Susceptibility Testing; S, susceptible; I, intermediate; R, resistant.

Results. In the last three years, 70 Acinetobacter baumannii were isolated from blood samples, and 62 were carbapenem-resistant. 62 CRAB strains were isolated from 62 patients using conventional culture media and tested for susceptibility to CFDC.

The range of diameters and the percentage of susceptible and resistant isolates are shown in **Table 1**, using interpretations of breakpoints recommended by EUCAST and CLSI. CRAB isolates showed higher susceptibility when CLSI breakpoints were applied compared to the EUCAST breakpoints. In particular, seven isolates with a zone diameter ≥ 15 mm were susceptible according to CLSI guidelines but resistant according to EUCAST breakpoints. When EUCAST breakpoints were applied, the overall susceptibility rate to CFDC was (45/62) 72.5%, whereas it was (52/62) 83.8% using CLSI breakpoints. Antibiotic susceptibility results for amikacin, colistin, and trimethoprim/ sulfamethoxazole of CRAB isolates according to susceptibility pattern to CFDC based on EUCAST breakpoints are shown in Table 2.

Discussion. In the present study, we tested the susceptibility of CRAB isolates to CFDC by using the Kirby-Bauer disc diffusion test on regular unsupplemented Mueller-Hinton agar, considered the gold standard method for this purpose. Then we compared the results to interpretation breakpoints recommended by EUCAST and CLSI guidelines.^{10,11} According to EUCAST breakpoints, in our study, resistance to CFDC was observed in 17/62 (27.4%) isolates, whereas by using CLSI guidelines, 10/62 (16.1%) isolates resulted in being resistant to CFDC. In previous epidemiological

studies investigating *in vitro* efficacy of CFDC against CRAB isolates, the overall rates of resistance ranged from 3.1% to 47.4%.¹²⁻²⁰ Therefore, the present results were within the range of literature data, although the range of resistance rates in previous studies was very wide.¹²⁻²⁰

Furthermore, susceptibility results were interpreted according to breakpoints recommended by EUCAST guidelines in some studies^{13,15,16,19} and by CLSI guidelines in others.¹²⁻¹⁷ In two of these studies, a disk diffusion test according to EUCAST breakpoints was used to assess the susceptibility of CRAB strains to CFDC, and the resistance rates to CFDC were $5.3\%^{14}$ and 22.1%,¹⁶ respectively. Compared to these studies, we found a higher resistance rate to CFDC among CRAB isolates. However, the study of Carcione et al., although conducted in Italy and during a period similar to ours, was based on a total of 19 isolates, and the small sample analyzed could at least partially explain the difference in resistance rate using DD (5%; 1/20) compared to our study.¹⁴ Conversely, the study of Ghebremedhin et al. was conducted in a different country (Germany) from 2014 to 2021, so their results could be difficultly comparable to ours.¹⁶

Furthermore, Morris et al. performed DD tests for CFDC by using 30-µg discs produced by two different manufacturers (i.e., FDA-cleared HardyDisks and MASTDISCS [RUO]) on standard MH agar on 14 CRAB strains and compared results by interpreting them according to both EUCAST and CLSI breakpoints; they found rates of resistance which were widely different depending on the type of disc used and interpretation breakpoints used: 35.7% and 57.1% by using

 Table 2. Antimicrobial susceptibility tests' results of carbapenem-resistant Acinetobacter baumannii strains to other choice antibiotics according to susceptibility pattern to cefiderocol based on EUCAST breakpoints.

			Carbapen	em-resistar	nt Acinetoba	cter bauman	<i>nii</i> isolates	(N=62)			
					Cefider	ocol					
Susceptible, % (N)					Resistant, % (N)						
72.5 (45)					27.5 (17)						
AN		CS		SXT		AN		CS		SXT	
S	R	S	R	S	R	S	R	S	R	S	R
% (N)	% (N)	% (N)	% (N)	% (N)	% (N)	% (N)	% (N)	% (N)	% (N)	% (N)	% (N)
9 (4)	91 (41)	100 (45)	0	9 (4)	91 (41)	70 (12)	30 (5)	94 (16)	6(1)	0	100 (17)

Abbreviations: S, susceptible; R resistant; AN, amikacin; CS, colistin; STX, trimethoprim/sulfamethoxazole.

HardyDisks discs and 7.2% and 64.7% by using MASTDISCS (RUO) discs, according to CLSI and EUCAST breakpoints, respectively; therefore, authors concluded that DD methods (at least with the methodology used in their study) performed poorly for CRAB.¹⁷ Based on these findings, it is possible to speculate that it is difficult to compare our results to those reported in the literature due to the heterogeneity of techniques used to assess the resistance of CRAB to CFDC and different local epidemiology or study periods. Because high rates of clinical failure were reported in patients affected by CFDC-resistant Grambacteria,²⁰ negative optimizing microbiological procedures to assess resistance as part of routine clinical practice is a mandatory task that should be the object of further investigations.

This study is affected by several limitations: i) this is a monocentric study, and the size of samples analysed is relatively small; ii) previous therapy with CFDC was not evaluated for patients carrying CFDC-resistant CRAB isolates; iii) analysis of the molecular characterization of resistance mechanisms was not conducted; iv) clonal analysis of CRAB isolates to identify possible local outbreak was not performed; this last limitation may have influenced the resistance rate reported in our study.

Conclusions. Our results showed a relatively high resistance rate to CFDC among clinical CRAB isolates compared to previous reports. However, several differences in methods, breakpoints interpretation guidelines, and local epidemiology should be considered.

Author Statement. Conceptualization, EMT and AQ; methodology, NM and AR; software, CC GGMS; investigation, VS, FS, RL, and VLB data curation, FD and GP; writing—original draft preparation, EMT and AQ; writing, EMT and AQ; supervision, CT and GM.

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Competing interests: The authors declare no conflict of Interest.

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