

Letter to the Editor**Fusion Gene Landscape in a Case of Acute Myelocytic Leukemia with Myelocyte Morphology****Keywords:** Fusion gene; Acute myelocytic leukemia; Myelocyte.**Published:** January 01, 2025**Received:** October 13, 2024**Accepted:** December 14, 2024**Citation:** Su Z., Li Y., Zhu H., Wang Y., Yang M. Fusion gene landscape in a case of acute myelocytic leukemia with myelocyte morphology. *Mediterr J Hematol Infect Dis* 2025, 17(1): e2025004, DOI: <http://dx.doi.org/10.4084/MJHD.2025.004>This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by-nc/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.**To the editor.**

A subset of acute myeloid leukemia (AML) exhibits leukemic cell morphology characterized by abnormal myelocytes. Such cases are primarily associated with the t(8;21)(q22;q22.1) chromosomal abnormality and the *RUNX1::RUNX1T1* fusion gene. The revised French-American-British (FAB) classification in China designates this as a new subtype, specifically M2b.¹ However, some M2b cases do not harbor the *RUNX1::RUNX1T1* fusion gene. Abnormal myelocytes are also sporadically observed in M4 and M6 cases according to the FAB classification. Limited information is available regarding the genetic features of these *RUNX1::RUNX1T1*-negative cases. In this study, we report an intriguing case of AML with leukemic cytomorphology characterized by abnormal myelocytes and monocytes. Transcriptome sequencing was employed to elucidate its fusion gene landscape.

The patient was admitted to our hospital for the first time due to "leukocytosis found for 2 days." Routine

blood tests indicated a white blood cell count of $23.08 \times 10^9/L$, neutrophil count of $9.65 \times 10^9/L$, monocyte count of $8.65 \times 10^9/L$, lymphocyte count of $4.74 \times 10^9/L$, hemoglobin level of 90 g/L, and platelet count of $19 \times 10^9/L$. Bone marrow smears showed that the proportions of myeloblasts, promyelocytes, abnormal neutrophils, and promonocytes were 4.5%, 2%, 28.5%, and 6%, respectively. Late erythroblasts with binucleated or petal-like nuclei were identified (**Figure 1A**). Blood smears revealed 15% myeloblasts, 12% myelocytes, 3% promonocytes, and 22% monocytes. Flow cytometry indicated that myeloblasts accounted for 21.54% of nuclear cells and expressed CD34, CD117, CD38, CD33, CD13, CD123, and HLA-DR. Promonocytes constituted 16.67% and exhibited the phenotypes CD38, CD64, CD123, CD33, CD13, CD36, and HLA-DR, with weak expression of CD11b. The results of chromosome karyotype analysis were normal (**Figure 1B**).

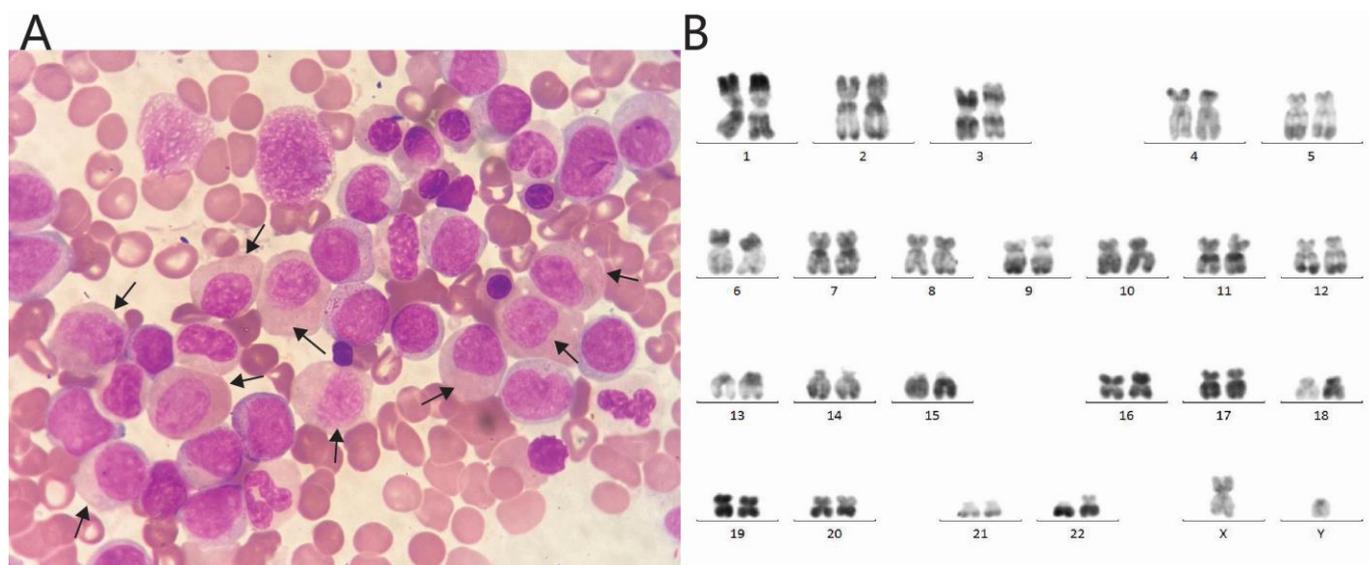


Figure 1. Cytomorphology and chromosome karyotype characteristics of the present case. (A) Blasts in bone marrow smear (1000 \times , Wright staining). Myelocytes are marked with arrows. (B) G-banded karyotyping.

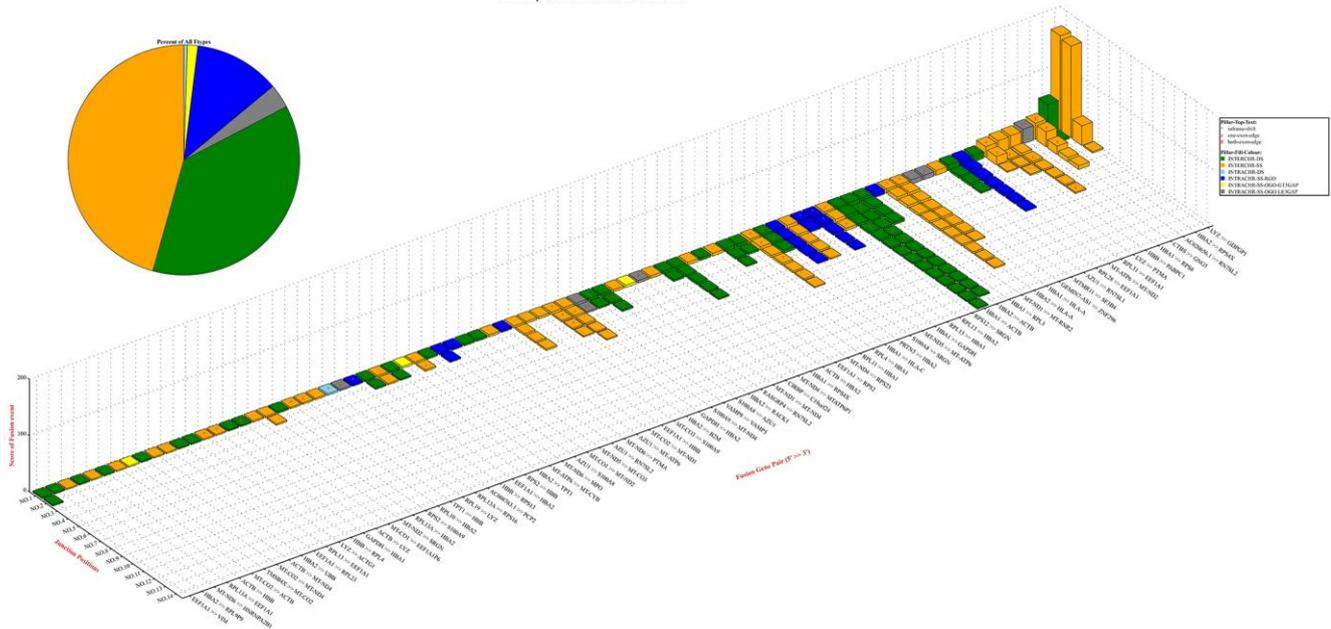


Figure 2. Fusion landscape of the present case.

Genetic mutation testing revealed *FLT3*-ITD (5.2%), *KRAS* c.38G>A (6%), *SRSF2* c.284C>A (47.8%), *ASXL1* c.2309C>A (47.4%), *RUNX1* c.941_942dup (47.4%), *STAG2* c.1821+2T>C (94.7%), *KMT2D* c.15671C>A (47.9%), and *TET2* c.2604T>G (47.6%). The diagnosis was acute myeloid leukemia (AML). The patient received a regimen of azacitidine (140 mg d1-7) and venetoclax (100 mg d1, 200 mg d2, 400 mg d3-14) for 2 courses, none of which achieved remission. The patient was eventually lost to follow-up.

Total RNA was extracted from the bone marrow mononuclear cells. Transcriptome sequencing was performed using the Illumina HiSeq 2500 instrument (Illumina, San Diego, CA). Employing SOAPfuse software, a series of fusion events was predicted (**Figure 2**).

In the 1950s, Yang and Yan et al. in China defined a unique form of AML. The predominant leukemic cells in the bone marrow are abnormal myelocytes. Compared with other AML leukemic cells, these abnormal myelocytes are larger, exhibit a relatively lower nuclear-to-cytoplasmic ratio, and show asynchronous development of the nucleus and cytoplasm. The nucleus contains fine chromatin dotted with one or two nucleoli. The abundant cytoplasm is typically basophilic. In the invagination of the nucleus, a homogeneous salmon coloration can be observed. The majority of these cells express stem/progenitor cell markers. Most reports suggest a high expression of CD34 and HLA-DR. CD15 is relatively highly expressed, whereas CD33 and CD13 are expressed at low levels, and CD7 is rarely expressed. The results of the literature review indicate that 40-98.2% of M2b

blasts express CD19, and 20-71.4% express CD56. According to China's revised FAB classification of leukemia, this form is classified as the M2b subtype. Most cases of M2b harbor the *RUNX1::RUNX1T1* fusion gene and the t(8;21)(q22;q22.1) translocation. Morphological features of M2b are also observed in other myeloid tumors, such as the acute phase of chronic myeloid leukemia, myelodysplastic syndromes, M4, and M6. However, the *RUNX1::RUNX1T1* fusion gene is absent in all of these cases.¹⁻³

In the case we present, cell morphology demonstrated a predominance of abnormal myelocytes (28.5%), while typical myeloblasts were rare (4.5%). The percentage of myeloblasts detected by flow cytometry was 21.54%, and these cells expressed stem/progenitor cell markers. Interestingly, the monocyte subpopulation exhibited similar characteristics, with cell morphology showing more mature features compared to the results from flow cytometry.

RNA sequencing (RNA-seq) revealed a series of fusion events, notably the absence of *RUNX1::RUNX1T1* and other frequent recurrent fusion genes. There are significant advantages of interchromosomal fusion compared with intrachromosomal fusion, as both coding and non-coding genes are involved. Some known oncogenic genes, such as *ZNF296* and *SF3B4*, can be identified as fusion partners.^{4,5} These partner genes are involved in various aspects of cellular function, including the regulation of transcription, pre-mRNA splicing, mRNA stabilization, tRNA delivery, protein synthesis, and degradation, the insertion of secretory proteins, glucose

metabolic processes, carbohydrate metabolism, ribosome construction, enzyme activity facilitation, inflammatory mediation, cell motility, cytoskeleton maintenance, apoptosis, and cell division. Multiple fusion genes are common in tumors and have different potential contributions to cancer development.⁶

In summary, we report a case of AML characterized by abnormal myelocyte and monocyte morphology in which the fusion gene landscape was delineated. Further cases are needed to elucidate the genomic features associated with this subtype.

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Competing interests: The authors declare no conflict of Interest.

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References:

1. Yushu H, Shougeng B, Zhijian X, et al. (1999) Acute myeloid leukemia M2b. *Haematologica* 84:193-194
2. Xiao Z, Hao Y, Bian S (1997) Acute myeloid leukemia M2B (subacute myeloid leukemia) in China. *Leukemia Res* 21:351-352. [https://doi.org/10.1016/S0145-2126\(96\)00101-4](https://doi.org/10.1016/S0145-2126(96)00101-4) PMID:9150353
3. 杨崇礼, 张新伟, 肖志坚, et al (2004) 急性髓系白血病M2b的研究进展. 白血病, 淋巴瘤 13:9
4. Mizoue Y, Ikeda T, Ikegami T, et al. (2023) The stem cell transcription factor ZFP296 transforms NIH3T3 cells and promotes anchorage-independent growth of cancer cells. *Int J Dev Biol* 67:147-153. <https://doi.org/10.1387/ijdb.230143hk>
5. Kidogami S, Iguchi T, Sato K, et al. (2020) SF3B4 Plays an Oncogenic Role in Esophageal Squamous Cell Carcinoma. *Anticancer Res* 40:2941-2946. <https://doi.org/10.21873/anticancerres.14272> PMID:32366446
6. Stephens PJ, McBride DJ, Lin M-L, et al. (2009) Complex landscapes of somatic rearrangement in human breast cancer genomes. *Nature* 462:1005-1010. <https://doi.org/10.1038/nature08645> PMID:20033038 PMCID:PMC3398135